

Effect of Organic Particulate Matter on Vegetable Crops and their Control Potentials

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Abstract. Air pollution has an impact, include impact to the plants. The adverse effects of air pollution have been associated with three major sources. They are gaseous pollutant and particulates from fossil fuels, manufacturing plants and vehicles. Many studies reveal that plants absorb particulate pollutants can cause damage or decline in growth. This study analysed the effects of organic particulate matter exposure on vegetable crops that were lettuce (*Lactuca sativa* L) and cucumber (*Cucumis sativus* L). Thus from the research we can control vegetable crops from particulate matter pollution. Growth parameters include the number of leaves, plant height, fresh weight, dry weight, number of chlorophyll and stomata index. This research was conducted on Padjadjaran University Greenhouse, Jatinangor. Lettuce and cucumber were given three treatment, those were: plant was contaminated with organic particulate matter derived from clay, plant inside and outside the chamber were not contaminated. The results showed that organic particulate matter exposure more affects the growth of lettuce than cucumbers for the following parameters: plant height, dry weight and fresh weight. Control potentials of organic particulate matter include regulation management, reducing field burning of vegetable residue, and implementation of particulate matter control devices in industry and vehicles.

Keywords: **Air Pollution; Controlling; PM₁₀ ; Vegetable Crops.**

1 Introduction

The increasing in population growth has results in increasing food demand. Previous researchers have studied food demand by Huang & Rozelle (1998), Abdulai, *et.al* (1999), Meenakshi & Ray (1999), Fuller, *et.al* (2000), Gould (2002), Ma *et.al* (2004), Wang and Zhou (2005), Rae (2008), Dong & Fuller (2010), Liu, *et.al* (2009), Gandhi and Zhou (2010), and Fu, *et.al* (2012) [1-12]. Regmi and Dyck (2001) have studied changes of food consumption across the countries [13]. And consumers are increasing their consumption on vegetables and fruits [14]. Global crop demand could increase from 5.5 to 10.9 Gton during 2011–2050 [15]. In the other hand, food productivity was threatened due to climate change. There is strong evidence that climate change affect food quality (diversity, nutrient density, and safety)[16-17]. This climate changes was caused by air pollution.

Air pollution has an impact, include impact to the plants. The adverse effects of air pollution have been associated with three major sources. They are gaseous pollutant and particulates from fossil fuels, manufacturing plants and vehicles. Many studies reveal that plants

absorb particulate pollutants can cause damage or decline in growth [18-23].

In order to provide appropriate air pollution controlling in horticulture crops, we analysed the effects of organic particulate matter on vegetable crops that were lettuce (*Lactuca sativa* L) and cucumber (*Cucumis sativus* L). The objective of the research was to analyze the effects of exposure of organic particulate matter on growth of lettuce (*Lactuca sativa* L) and cucumber (*Cucumis sativus* L) both morphology and physiology thus from the research we can control vegetable crops from air pollution.

Air pollution controlling on vegetable crops is one of the solution to mitigate vegetable quality and production. Good quality of vegetable crops will directly stimulate healthier society thus may results in developing low carbon society.

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2 MATERIALS AND METHODS

2.1. Area of Study

This research was laboratory scale and was conducted on Padjadjaran University Greenhouse, Jatinangor, Sumedang District, West Java, Indonesia. It is located at 723 meters above sea level. The climate of the area is relatively temperate with an average rainfall of about 2500 mm. The highest temperature recorded was 19°C-27°C.



Fig. 1. Padjadjaran University Greenhouse

2.2 Experimental Setup

Lactuca sativa L and *Cucumis sativus* L were given three treatment, those were: plant was contaminated with organic particulate matter derived from clay, and plant inside and outside the chamber were not contaminated.

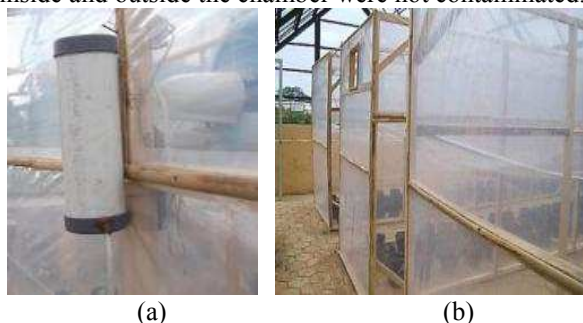


Fig. 1. Experimental Setup (a) clay (b) chamber design

2.3 Air Pollution Monitoring

Pollutant parameters measured in the chamber contaminated with organic particulate matter derived from clay was PM₁₀. PM₁₀ was monitored every 15 minute of 120 minute intervals for 4 (four) weeks.

2.4 Plant Sampling and Analysis

Plants were grown under field conditions in Green House. Plant samples were analysed every 7 days of intervals for 4 (four) weeks. Growth parameters include the number of leaves, plant height, fresh weight, dry weight, number of chlorophyll and stomata index.

ANOVA was used to determine the significant difference between treatments for the different variables and the Least Significant Difference (LSD) test at the 95%

probability level. All the statistical tests were performed using SPSS software (SPSS Inc., version 10).

2.4 Stomata Observation

Observation of leaf structure and stomata using light microscope with 400x enlargement and using Scanning Electron Microscope (SEM) was performed at Instrumentation Laboratory in ITB.

3 RESULTS

3.1 Concentration of Particulate Pollutants

The chamber was contaminated with clay, pollutant parameters measured was PM₁₀. The results of particulate concentrations are shown in Table 1. During the exposure period, particulate concentration in clay chamber tends to be higher than the measured control levels both control inside or outside. Maximum particulate concentration was found at second week (1.677,78 µg/m³).

Table 1. Particulate Concentration During Exposure Period

Treatment	PM ₁₀ Concentration (µg/m ³)			
	1st Week	2nd Week	3rd Week	4th Week
Control Outside	200,97	198,46	22,69	55,56
Control Inside	116,67	76,39	45,37	912,04
Clay Chamber (Particulate)	325,00	1.677,78	195,27	95,83

3.2 Growth Response

The results of growth response of PM₁₀ exposure to the plant are shown in Table 2 & Table 3 and Figure 3. Plant growth parameters were plant height, number of leaves, plant fresh weight and plant dry weight.

Table 1. Plant Morphology Parameters on *Lactuca Sativa* L Exposed To Pollutant

Plant Samples	Control Outside	Control Inside	Particulate Pollutant
PH	35.4167a	44.6667b	40,0417c
NL	19.1250a	21.9583c	21,5417bc
PFW	135,9233a	116,1994b	105,4480b
PDW	12,3829a	10,1888b	9,5594b

Note: y = Means within columns having different letters are significantly different according to the least significant difference (LSD) at 0.05 level of probability. PH: Plant Height, NL: Number of Leaves, PFW: Plant Fresh Weight, PDW: Plant Dry Weight.

Treatment of particulate exposure was significant (P < 0.05) on decrease of plant height of *Lactuca sativa* L compared to control inside rather than control outside (Sig 0.000 < 0.05). Meanwhile there was no significant

difference on number of leaves from four treatments. Particulate exposure was significant ($P < 0.05$) on decrease of plant fresh and dry weight of *Lactuca sativa* L compared to control outside rather than control inside (Sig $0.000 < 0.05$).

Table 3. Plant Morphology Parameters on *Cucumis Sativus* Exposed To Pollutant

Plant Samples	Control Outside	Control Inside	Particulate Pollutant
PH	118,9167a	129,6625b	132,1875b
NL	13.4583a	25,9167b	27,4167b
PFW	56,6351a	69,3773a	70,9403a
PDW	4,5298a	6,1328b	6,9918b

Note: y = Means within columns having different letters are significantly different according to the least significant difference (LSD) at 0.05 level of probability. PH:Plant Height, NL: Number of Leaves, PFW: Plant Fresh Weight, PDW: Plant Dry Weight.

Based on statistical study, there was no effect of particulate exposure on reduction on growth of *Cucumis sativus* L ($P < 0.05$).

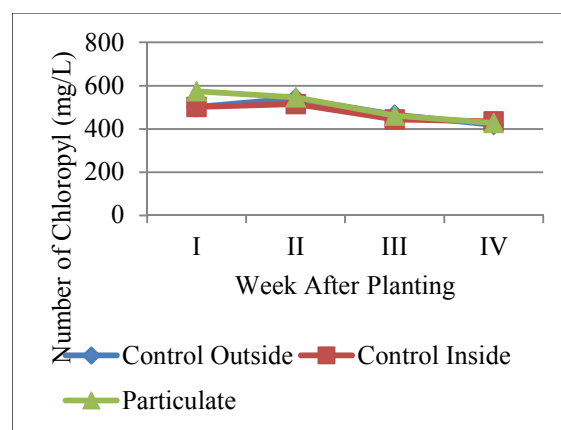


(a) (b) (c)

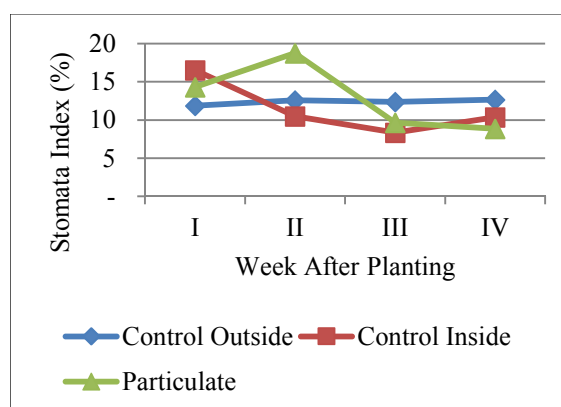
Fig. 3. Growth Responses of *Lactuca sativa* L to Particulate Exposure (a) control outside, (b) control inside, (c) Particulate exposure

3.2 Physiological Response

Physiological parameter was number of chlorophyll and stomata index. The results of physiological response of both plants to particulate exposure are shown in Figure 4 and Figure 5.

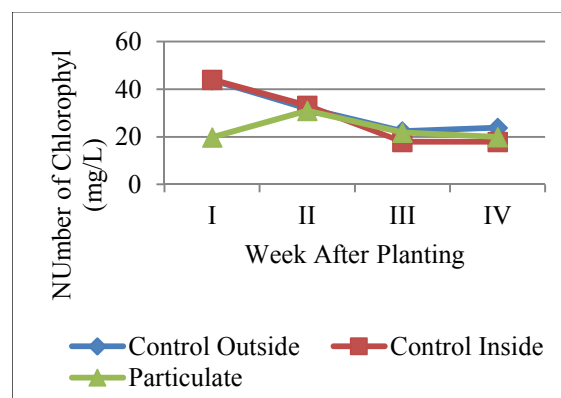


(a)

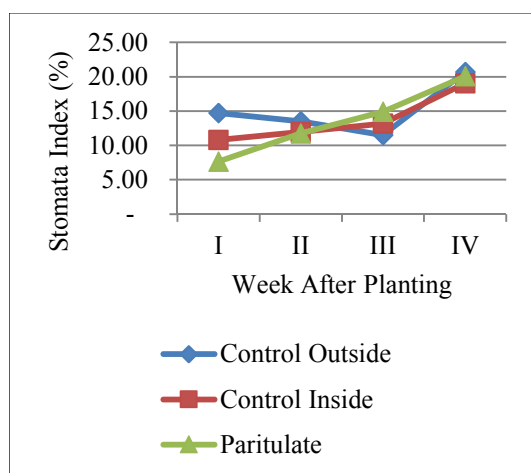


(b)

Fig. 4. Physiological Responses of *Lactuca sativa* L to Particulate Exposure



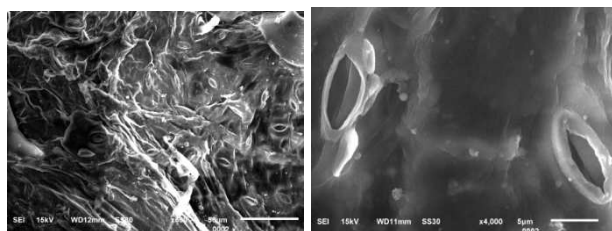
(a)



(b)

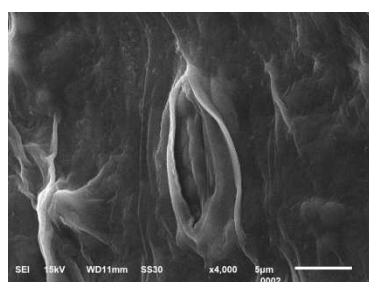
Fig. 5. Physiological Responses of *Cucumis sativus* L to Particulate Exposure

According to the study, there were no effect of particulate exposure on reduction on number of chlorophyll and stomata index of both plants during the particulate exposure period compared to control outside and control inside chamber.



(a)

(b)



(c)

Fig. 6. Stomata of *Lactuca sativa* L under SEM Observation (a) Control outside (b) Control inside (c) Particulate Exposure

4 DISCUSSIONS

Based on study, particulate exposure was significant ($P < 0.05$) on decrease of plant height, plant fresh and dry weight of *Lactuca sativa* L compared to control outside rather than control inside ($Sig\ 0.000 < 0.05$).

Previous studies have been done on pollutant for damaging effect to the plant. The main responses are morphological and on the development of flowers and fruit [24]. Another study concludes that excess substance

become toxic to the plant. This caused leaf chlorosis and root browning. Greater toxic substance concentration in the nutrient solution along with the appearance of toxicity symptoms significantly depressed the fresh mass of leaves, stems and roots [25]. Phosphate and nitrogen are plant nutrient in small quantities; excess nutrient will damage the plant. There is a complex change in physiological response due to excess substance. Plant cell may cause lysis and damage. Naama et.al on their study on fungal spores showed the allegenicity since the fungal spores were exposed to air pollution. This is due to protein nitration and deamidation [26].

Plants are the primary receptors for pollutants in the atmosphere including particulate matter. This is due to huge foliar surface area in the upper epidermis of the leaves that acts as natural sink for pollutant. The harmful effects of pollutant especially particulate matter on vegetation have already been note by many researchers [27-36]. From previous study, it showed that vegetation is an effective indicator of impact of air pollution especially particulate matter.

Based on study, the particulate exposure more affects the morphology of *Lactuca sativa* L than *Cucumis sativus* L. Impact of pollutant depends on the concentrations of elements in the chamber and the physiological status of the plant.

Previous study showed that phytotoxicity level of plant due to environmental contamination as follows: *Z. mays* < *C. sativus* < *L. sativa* L. The results indicate inhibition in root elongation as the most sensitive toxicity end point for *L. sativa* L [37]. This study conclude that *C. sativus* L was tolerant vegetable to particulate pollutant than *L. sativa* L. The studies of the responses of species to air pollutants for their tolerance or sensitivity has been done by Gao et al. (2016); Mukherjee and Agrawal, (2016); Singh et al., 1991; and Wen et al.,(2004) [38-41].

Based on study, we may conclude that there was effect of the particulate pollutant to vegetable crops especially on the sensitive vegetables. Vegetable planting should consider air pollution controlling in order to maintain and enhance vegetable productivity. In Indonesia, air pollutants are the products of combustion from industrial area and transportation sectors which is currently developing. Olivier et.al (2016) reported that Indonesia (currently with a share of 1.4% of the global total CO₂ emissions) showed a 4.0% increase in CO₂ emissions in 2015, compared to 2014. These CO₂ emissions derived from power and heat generation, other energy industry own use, manufacturing industry, road transport, other transport, residential sector, and other buildings [42]. Therefore, air pollutant controlling on vegetable crops include setting vegetable crops land away from industrial area and transportation sectors.

In Indonesia, the farmers used to burn crops residue after harvesting, therefore it may increase air pollutant exposure. Agricultural activities are the major human source of air pollution in rural areas. Majra (2011) stated that burning of stubble in the field after harvesting,

threshing operation, grain dust and large scale use of tractors harvester, combines and diesels operated tube well are major factor contributing to air pollution [43]. According to Satyendra et.al (2013), burning of these residues emit gases like sulphur dioxide (SO₂), oxides of nitrogen (NO_x), carbon dioxide (CO₂), carbon monoxide (CO), black carbon (BC), organic carbon (OC), methane (CH₄), volatile organic compounds (VOC), non-methane hydrocarbons (NMHCs), ozone (O₃), and aerosols which affect the global atmospheric [44]. Burning of crop residues emitted 8.57 Mt of CO, 141.15 Mt of CO₂, 0.037 Mt of SO_x, 0.23 Mt of NO_x, 0.12 Mt of NH₃ and 1.46 Mt NMVOC, 0.65 Mt of NMHC, 1.21 Mt of particulate matter for the year 2008–09 in India [45]. These air pollutants may distribute in the atmosphere and may affect the crops planting on the other area. Based on fact, the air pollutant controlling on vegetable crops was avoiding open burning after harvesting. According to Satyendra et.al (2013), burning of crop residue/biomass can be avoided by adopting different biochemically/thermo-chemically induced techniques. Technologies are available for harnessing energy from crop residues are direct combustion, gasification, carbonisation, ethanol production, liquefaction, bricking and pyrolysis [44]. Other researchers suggest using agricultural residues as feedstock for biofuel production [46-47].

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References

- J. Huang, S. Rozelle, China Eco. Re. **9**, 25–45 (1998)
- A. Abdulai, D. K. Jain, A. K. Sharma, J.of Agr. Eco. **50**, 316–327 (1999)
- J. V. Meenakshi, R. Ray, R, J.of Intl. Dev.**11**, 47–74 (1999)
- F. Fuller, D. Hayes, D. Smith, Eco. Dev. & Cultl. Change **49**, 23–43 (2000)
- B. W. Gould, Agr. **18**, 387–407 (2002)
- H. Ma, J. Huang, S. Rozelle, Eco. Dev.and Cultl. Change **52**, 445–473 (2004)
- J. M. Wang, Z. Y. Zhou, Aldershot:Ashgate 87–107 (2005)
- A. Rae, Aus. Jour. of Agr. and Re. Eco. **52**, 283–302 (2008)
- F. Dong, F. Fuller, Can. Jour. of Agr. Eco. Rev. Can. d'agroeco. **58**, 73–91 (2010)
- H. Liu, K. A. Parton, Z. Y Zhou, R. Cox, Aus. Jour. of Agr. and Re. Eco. **53**,1 (2009)
- Gandhi, P. Vasant Z. Y Zhou, Aus. Agr. Rev. **18**, 103–135 (2010)
- Fu, Wenge, Gandhi, P. Vasant, Cao, Lijuan, Liu, Hongbo, Zhou, Zhangyue, Chi. & Wrld Eco. **20**, 88–106 (2012)
- A. Regmi, J. Dyck, *Effects of urbanization on global food demand. In A. Regmi (Ed.), Changing structures of global food consumption and trade.* (ERS WRS 01-1, Economic Research Service, Washington, DC: United States Department of Agriculture 2001)
- P. Vasant, Gandhi,Z. Zhangyue, Food Resour. Intl. **10**, 963 (2014)
- GLOPAN (Global Panel on Agriculture and Food Systems for Nutrition), *Climate smart food systems for enhanced nutrition* (London, 2015)
- S. J. Vermeulen, B. M. Campbell, J. S. I. Ingram, Annu. Rev. Environ. Resour. **37**, 195–222 (2012)
- S. K. Prajapati, Environ.Skept.Critics **1**,12–22 (2012)
- U. Younis, T. Z. Bokhari, S. A. Malik, S. Ahmad, R. Raja, Int.J.Agric.Sci.Res.**3**,1–12 (2013)
- P. K. Rai, L.S. Panda, B. Chutia, M. Singh, Afr.J.Environ.Sci. Technol.**7**, 944–948 (2013)
- P. K. Rai, L. S. Panda, AirQual.Atmos.Health **7**, 93–101 (2014)
- P. K. Rai, M .M. Singh, J.Asia-Pac.Biodivers. **8**, 375 (2015)
- P. K. Rai, *Biomagnetic Monitoring through Roadside Plants of an Indo-Burma Hot Spot Region* (Elsevier,UK, 2016)
- P. K. Rai, J. Asia-Pac. Biodivers. **9**, 47–55 (2016)
- J. M. Wang, Z. Y. Zhou, Aldershot:Ashgate 87–107 (2005)
- A. Rae, Aus. Jour. of Agr. and Re. Eco. **52**, 283–302 (2008)
- F. Dong, F. Fuller, Can. Jour. of Agr. Eco. Rev. Can. d'agroeco. **58**, 73–91 (2010)
- R. Misra, P. K. Behera, Pollut.Res. **13**,203–206 (1994)
- M. Farooq, K. R. Arya, S. Kumar, K. Gopal, P/ C. Joshi, R. K. Hans, J. Environ.Biol. **21**,165–167 (2000)
- N. Shrivastava, S. Joshi, Geobios **29**,281–282 (2002)
- I.N. Gostin, Not. Bot. Horti. Agrobot. Cluj-Napoca **37**, 57–63 (2009)
- D. Sukumaran, *Effect of Particulate Pollution on various Tissue Systems of Tropical Plants* (Central Pollution Control Board (CPCB), Zonal Office, Kolkata, India, 2012)
- S. S. Garg, N. Kumar, G. Das, Indian J. Environ. Prot. **20**,326–328 (2000)
- J. Garty, O. Tamir, I. Hassid, A. Eshel, Y. Cohen,A. Karnieli, J. Environ. Qual. **30**,884–893 (2001)
- P. M. Mashitha, V. I. Pise, Pollut. Res. **20**,195–197 (2001)
- J. G. Gavali, D. Saha,K. Krishnayya, Indian J. Environ. Health **44**, 88–91 (2002)
- P. K. Rai, Atmos. En-viron. **72**,113–129 (2013)
- K. Masakorala, J. Yao, H. Guo, R. Chandankere, J. Wang, M. Cai, H. Liu, M. M. F. Choi, W. A. S. Poll **224**, 1553 (2013)
- F. Gao, V. Calatayud, F. Garcia-Breijo, J. Reig-Armiñana, Z. Feng, Ecol. Indic. **67**, 367 (2016)
- A. Mukherjee, M. Agrawal, Bull. Environ. Contam. Toxicol. **96**, 1-6 (2016)
- S.K. Singh, D.N Rao, M. Agrawal, J. Pandey, D. Naryan, J. Environ. Manag. **32**, 45–55 (1991)
- D. Wen, Y. Kuang, G. Zhou, Environ. Sci. Pollut. Res. Int. **11**, 165–170 (2004)
- A. Mukherjee, M. Agrawal, Eco. E. S **152**,42 (2018)

43. J.G.J. Olivier, G. Janssens-Maenhout, M. Muntean, J. A.H.W. Peters, *Trends in Global CO2 emissions: 2016 report* (PBL Publishers, European Commission, 2016)
44. J.P. Majra, *Air Quality in Rural Areas* (INTECH, 2011)
45. T. Satyendra, R.N. Singh, S. Shaishav, *Int. R. J. E. Sci* **1**, 24 (2013)
46. N. Jain, A. Bhatia, H. Pathak, *Aer.A.Q.R* **14**, 422 (2014)
47. T. Searchinger, R. Heimlich, R. A. Houghton, F. Dong, A. Elobeid, J. Fabiosa, S. Tokgoz, D. Hayes, T. H. Yu, *Sci*, **319**, 1238–1240 (2008)

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