

Understory and Soil Macrofauna Diversity under the Three Young Native Species and *Acacia crassicarpa* in a Drained Peatland of Pelalawan-Riau, Indonesia

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Abstract. Most studies mentioned that *Acacia crassicarpa* belongs to invasive species that could threaten the native biodiversity. To respond to that issue, we conducted a study that covers the understory and soil macrofauna diversity of three native tree species, namely mahang (*Macaranga pruinosa*), skubung (*Macaranga gigantea*) and geronggang (*Cratoxylum arborescens*) and an exotic species namely krassikarpa in a drained peatland in Pelalawan, Riau. The observation of understory vegetation under each studied species was undertaken by using 2 x 1 m plots. Furthermore, the structure of macrofauna was observed by pitfall trap methods. Results revealed that there were two fern species namely *Neprolepis biserrata* and *Stenochlaena palustris* that dominated the understory vegetation in mahang (*Macaranga pruinosa*), skubung (*Macaranga gigantea*) and geronggang (*Cratoxylum arborescens*). The diversity index in vegetation structure among those four tree species was insignificantly varied. Moreover, the percentage of understory coverage under *A. crassicarpa* was significantly higher than that under all native tree species. On the other hand, Formicidae and Rhinotermitidae were dominant in skubung and krassikarpa. Meanwhile, Formicidae and Blattidae were high in mahang and geronggang. Furthermore, diversity index of macrofauna were significantly low at krassikarpa's understory compared to other three native species. This study suggested that the introduction of krassikarpa affects the biodiversity.

Keywords: Mahang, skubung, geronggang, krassikarpa, understory and soil macrofauna

I Introduction

Indonesia is the second largest biodiversity country in the world after Brazil [1,2]. In order to maintain its biodiversity richness, Indonesia established 566 national parks that almost protect 36 million ha [3]. Furthermore, Indonesia has at least 88 million ha, including tropical peat swamp forests that are about 21 million ha of this type of tropical forest in Kalimantan, and Papua were disturbed [4,5]. Peat is one of soil types which has more than 18% of C-content and commonly found in one ecosystem located behind the riverbank [5,6]. One characteristic of peatland is that the land is easily and rapidly loses its water content and it leads to desiccation [7]. When the desiccation is widely and heavily spreading, it is easily being burned and would lead to a more disastrous situation.

Besides its vulnerable characteristics, biodiversity loss is the other problem which is more challenging in peatland. Forest degradation and conversion into agriculture purposes, unsustainable and irreversible forest management, expanding of infrastructure, slash and burn, and illegal mining, has been blamed for biodiversity loss in peatland [5,8–11]. Yet, another challenge is coming from the introduction of alien species that could threaten our native

species and biodiversity. There are many studies mentioned about existence of *Acacia* species which is sometimes called as invasive species that was harmful for native species since it has characteristic as invasive species [12–15]. In Riau province, one of exotic *Acacia* species i.e. *A. crassicarpa* (krassikarpa) has been commonly grown in peatland by two biggest pulp and paper industries (APRIL and APP Sinar Mas). In addition, *A. crassicarpa* was originally distributed in Australia and south Papua [16].

The existence of *A. crassicarpa* in Riau province gives positive impacts in social and economy side. Nonetheless, there is no many studies that concern in biodiversity structure changing formed by the existence of *A. crassicarpa*. Thus, study that reveals the effect of *A. crassicarpa*'s existence needs to be conducted. The most common methods to answer the questions is by doing a comparative study that comparing macrofauna and understory vegetation of the exotic tree species (*A. crassicarpa*) and local native trees species (*M. gigantea*, *M. pruinosa*, and *C. arborescens*).

Understory vegetation is one of biodiversity group that not only have responsibility in soil and water conservation but in erosion preventing also [17,18]. Furthermore, according to Lisnawati et al. [6] soil fauna holds the important role in restoring physics, chemistry, and biology structure through immobilization and humification. As a result, their existence has been considered as soil quality bioindicator [6,19]. Hence, based on those problems and issues, the objective needs to be addressed in this study was to examine the structure and diversity of understory and macrofauna between exotic tree species and three native trees species.

2 Material and methods

2.1 Study site and time

This study was conducted in a drained peatland at Lubuk Ogong, Pelalawan regency, Riau province (101°41'06"–101°41'10" E, 0°19'42"–0°19'48" N and 12 m asl) during March 2014 to December 2014. The peat type at location based on its maturity is dominated by fibric-hemic and drained with the annual variation of water table depth was 20 – 140 cm below soil surface [20,21]. The plots were consisted of four years old of *A. crassicarpa*, *M. gigantea*, *M. pruinosa*, and *C. arborescens*.

2.2 Design of observations

In the beginning, the plots were established in 2011 by using Randomized Complete Block Design (RCBD) thus it was three years old already when this study was conducted. The plots were originally designed for silviculture intensive treatments. There were four tree species planted in the plots and each tree species had five replications. Every replication had 400 m² and been planted with 3 x 2 m of those four tree species.

2.3 Sample collections

Understory samples were collected by arranging five rectangles of 2 x 1 m that were set in every plot of tree species (*A. crassicarpa*, *M. gigantea*, *M. pruinosa*, and *C. arborescens*). Plots were determined by systemic random sampling following diagonal formation of plots' size. Moreover, five replications were conducted for each species. Thus, we had 25 plots for each tree species. Vegetations were analyzed up to species level.

Macrofauna samples were collected by pitfall trap method. There were three traps that were set for each tree species. The traps were determined by systemic random sampling following diagonal formation of plots' size so that we had 15 traps for each tree species. Macrofauna were analyzed up to family level based on book Study of Insects written by Charles A. Triplehorn, Norman F. Johnson, and Donald Joyce Borror.

2.4 Data preparation and analysis

The collected data was tabulated systematically in order to obtain the result of Important Value Index (IVI) of the understory vegetations and macrofauna. This data is important to determine the most dominant species that available in the plots. IVI was approached by determining the value of relative frequency (RF) percentage that was added by relative density (RD) percentage. Furthermore, the value of diversity index was determined using Shanon-Wiener equation (Odum, 1993).

$$H = - \sum \{(n.i/N) \ln (n.i/N)\}$$

H' = diversity index

n.i = important value of N species

N = total of important value

Furthermore, the data of IVI of understory and macrofauna was analyzed by descriptive quantitative to figure out the most dominant species that occupied the understory and its structure. Meanwhile, anova was used to determine the tree species that had most significant value of understory and macrofauna diversity index. Tukey test was performed when anova's result showed any significant results.

3 Results and discussions

3.1 Understory vegetation

In general, number and diversity species within a community describe a picture of the richness and diversity of its community. Meanwhile, species domination is the sign of special entity of the community. The results revealed that ferns species, namely *Nephrolepis biserrata* and *Stenochlaena palustris* dominated the understory vegetation understands of *M. gigantea*, *M. pruinosa*, and *C. arborescens*. Yet, *N. biserrata* was more dominant than *S. palustris* in all three native species (Table 1). Similarly, in exotic tree species (*A. crassicarpa*), *N. biserrata* and *S. palustris* became the most dominant vegetation that occupied *A. crassicarpa*'s understory. Moreover, *N. biserrata* had four times of IVI than *S. palustris* (Table 1). Interestingly, the second largest group that dominated the three native tree species and the exotic species were varied. In *M. gigantea* and *M. pruinosa*, narrow leaf weed is dominant than broad leaf weed. Even *Scleria sumatrensis* and *Ottochloa spinosa* were two species found dominant after ferns in *M. gigantea*. In contrast, *C. arborescens* did not have any narrow leaf weed in its understory and were dominantly occupied by broad leaf weed named *Melastoma malabatricum*. Meanwhile, seedlings and saplings of *A. crassicarpa* were dominant in its understory after ferns.

Table 1. IVI of understory's vegetation of *A. crassicarpa*, *M. gigantea*, *M. pruinosa*, and *C. arborescens*.

Species	Family	Important Value Index			
		<i>M. gigantea</i>	<i>M. pruinosa</i>	<i>C. arborescens</i>	<i>A. crassicarpa</i>
<i>Nephrolepis biserrata</i>	Nephrolepidaceae	25.39	25.99	26.34	52.05
<i>Stenochlaena palustris</i>	Blechnaceae	14.68	13.32	22.69	12.61
<i>Asystasia</i> sp.	Acanthaceae	3.94			3.73
<i>Melastoma malabatricum</i>	Melastomaceae	3.94	6.76	11.74	
<i>Scleria sumatrensis</i>	Cyperaceae	13.96	11.20		3.73
<i>Ottochloa spinosa</i>	Poaceae	4.65			
Seedlings and saplings of <i>Acacia crassicarpa</i>	Fabaceae	3.94			9.58
<i>Colopogonium</i> sp.	Fabaceae		8.88	3.67	
<i>Mikania micrantha</i>	Asteraceae		3.73		
<i>Smilax</i> sp.	Smilacaceae				3.73

Pteridophyta division was the vegetation that dominantly occupied the understory structure of under all trees species either in native or exotic trees species. *S. palustris* and *N. biserrata* were two species of ferns that dominated vegetation structure. Moreover, besides *Dicranopteris* sp. and *Blechnum orientale*, *S. palustris* and *N. biserrata* are two species of ferns that dominantly covered the floor of *A. crassincarpa* planted in peat land [22–24]. A study conducted by Harnelly et al. [24] revealed that *N. biserrata*, *B. orientale*, and *Pteris trimula* that are classified as fern, dominantly grew at the Rawa Tripa Peat Swamp Forest, Aceh. It means that those species were highly adaptive not only at native trees species but at exotic tree species also. Furthermore, *N. biserrata* had a tendency and prefer an area that was not inhabited by trees and rapidly cover post burned area [4,5]. In other words, *A. crassincarpa* had more level of light penetration compare to other native trees species. As a result, the IVI of *N. biserrata* at *A. crassincarpa* had IVI fourtimes was doubled than IVI of other native trees species. Thus, it is assumed that the condition under *A. crassincarpa* was suitable for *N. biserrata* to grow and develop.

The interesting facts occurred in the second group that dominated the floor of those exotic and native trees species. Narrow leaf group were dominantly occupied at *M. gigantea* and *M. pruinosa* that was represented by *S. sumatrensis* and *O. spinosa*. In contrast, *M. malabatrimum* that categorized as broad leaf group was the second dominant species at *C. arborescens*. Meanwhile, seedlings and saplings of *A. crassincarpa* were the second species that were highly found in *A. crassincarpa*'s floor after ferns. Narrow leaf group abundancies were high when area was exposed to light penetration, particularly in the first year of *A. crassincarpa* and significantly decline when age of *A. crassincarpa* incline [22]. Other studies mentioned that spacing and grass weed (narrow leaf) has ability to inhibit the development of broad leaf seeds [26,27]. Meanwhile, there is no narrow leaf group could be found at *C. arborescens*'s understory and even *M. malabatrimum* which is a broad leaf that dominated the floor. *C. arborescens* has wider and denser canopy than *M. pruinosa*, *M. gigantea*, and *A. crassincarpa*. It reveals that broad leaf are more resistant than narrow leaf in case of low light penetration. When this phenomenon is compared to *A. crassincarpa*'s understory that is occupied by seedlings and saplings of *A. crassincarpa*, we assumed that the condition formed by *A. crassincarpa* is only suitable for ferns and not suitable for other local or native weeds, such as *S. sumatrensis* and *M. malabatrimum*. A report already stated that *A. crassincarpa* belongs to invasive species even though it was restricted in certain location [28] and its sister, *Acacia mangium*, has been grouped into invasive species also [29]. Hence, seedlings and saplings of *A. crassincarpa* are the second most species that could determine the understory after ferns.

Furthermore, the density or percentage of understory coverage area was significantly high in *A. crassincarpa* than other native trees species ($p < 0.05$). Interestingly, the percentage of understory's coverage area in *A. crassincarpa* was 100%. In other words, the understory under *A. crassincarpa* was fully covered by vegetations which was dominantly condensed by ferns (Table 2). In contrast, the other three native trees species had a low percentage of understory's coverage area (<30%) (Table 2).

Table 2. Percentage of understory coverage area of *A. crassincarpa*, *M. gigantea*, *M. pruinosa*, and *C. arborescens*.

Species	Percentage of understory coverage area
<i>M. gigantea</i>	29.45 ± 6.73a
<i>M. pruinosa</i>	24.45 ± 13.47a
<i>C. arborescens</i>	27.77 ± 10.71a
<i>A. crassincarpa</i>	100b

Remark: the alphabets appear after the value showed a significant value ($p < 0.05$).

In addition, percentage of understory coverage was significantly high and 100% fully covered the floor of *A. crassincarpa* than any other native trees species. When this data was combined to IVI that placed ferns as the most dominant species, it could be assumed that the condition is appropriate for ferns growth and development. The first factor that could determine this phenomenon is the interaction between genetics and environment, such as soil, microclimate, microorganism, and competition with other species [9,10]. Secondly, the ability of the species to propagate themselves [10,11]. Moreover, most ferns grow well in 40%-50% of light penetration, 60%-80% of humidity (to inhibit desiccation), temperature of 21 °C-27 °C, and attach themselves into humid soil [32]. Moreover,

Kerb [33] stated that the capability of certain species to occupy an area is influenced not only by its ability in establishing the adaptation mechanism toward all abiotic factors, such as temperature, light, soil, humidity, etc. and biotic factors, such as intra species interaction, competition, parasitism, etc. but also chemistry factors that include water, oxygen, pH, and nutrition. Hence, it could be assumed that condition under *A. crassicalpa* is appropriate for ferns. In the opposite, the phenomenon at three native trees species that only coverage under 30% of its floor showed that the environment condition created by those three native trees species are not favorable and tend to inhibit the development and growth of understory vegetation.

Besides, the diversity index value of *A. crassicalpa* and other three native trees species revealed that there were no significant differences ($p > 0.05$). It means that the diversity index of those species was statistically similar (Table 3). Nonetheless, *A. crassicalpa* had the lowest diversity index value than the other all native tree species. Moreover, *M. gigantea* had the highest value of diversity index and three times of diversity index value of *A. crassicalpa*.

Table 3. Diversity index of understory's vegetation of *A. crassicalpa*, *M. gigantea*, *M. pruinosa*, and *C. arborescens*.

Species	Diversity index (shanon-weiner)
<i>M. gigantea</i>	$1.80 \pm 0.47a$
<i>M. pruinosa</i>	$1.13 \pm 1.03a$
<i>C. arborescens</i>	$0.98 \pm 0.31a$
<i>A. crassicalpa</i>	$0.61 \pm 0.08a$

Remark: the alphabets appear after the value showed a significant value ($p < 0.05$)

Diversity index analysis showed that there was no significant between three native trees species and exotic tree species ($p > 0.05$). Thus, it means that diversity index between all tree species is similar. Nonetheless, *A. crassicalpa*'s understory had the lowest diversity index than other three native trees species. Even though floor of *A. crassicalpa* was fully and dominantly covered by ferns, the diversity index was low. This results was similar to a study conducted by Bela et al. [34] that revealed that diversity index of an peat land area that were dominated by fern (*Dicranopteris linearis*) was 0.3. Hence, we assumed that ferns successes in occupying all the resources and do not give any opportunity for other seedlings and saplings to grow and develop. As a result, there was not many of seedling and sapling varieties that could compete and grow between ferns' domination. This assumption was supported by Jackson and Finley [35] that found at least two ways of ferns in inhibiting seedling germination, namely allelopathy and less light intensity (below 0.5 percent of full sunlight). Consequently, the seedlings and saplings of other species cannot be germinated and there was not much variation of species.

3.2 Macrofauna

The results revealed that Formicidae is family that dominated the floor of either exotic tree species or the three native tree species ferns species (Table 4). Compare to other families, Formicidae had five different groups than other families. Moreover, class of collembola was dominantly found in *A. crassicalpa* than other three native tree species observed. In addition to that, Rhinotermidae was the second largest family found in *A. crassicalpa* and *M. gigantea*. Meanwhile, bug family (Curculionidae) and Polydesmidae were the second largest family found in *M. pruinosa*. In *C. arborescens*, Blattidae was a family dominantly found after Formicidae. Interestingly, group of macrofauna that dominated all the four tree species either exotic tree species or native trees species belong to social insects, such as Formicidae and Rhinotermidae.

In macrofauna, Formicidae, which is represented by Formicidae, was the dominant family that occupied either exotic tree species or all three native trees species. Meanwhile, Rhinotermidae was the second largest family after Formicidae. Formicidae and termites are two groups of soil fauna that dominantly inhibit peat soil [36-38]. Formicidae are a group of insects that is commonly distributed almost in everyplace and have characteristics to adapt in varies types of peat soil [6]. Even, compare to other insects group, Formicidae are the most success group [39]. The ability of Formicidae to dominate ecosystem is supported by their characteristics as carnivore, saprophyte, predator, and

detritivore. Formicidae has potency as a bioindicator of peat soil quality since they are able to live in an area that has wider range of pH, soil humidity, and peat land types. Interestingly, *A. crassicarpa* had only two groups of Formicidae than other native trees species. According to Lisnawati et. al. [6], the number of formicidae describes soil humidity since Formicidae prefers to live in soil that have low in humidity. Hence, it can be inferred that soil humidity is higher in *A. crassicarpa* than other native trees species.

Table 4. IVI of soil macrofauna of *A. crassicarpa*, *M. gigantea*, *M. pruinosa*, and *C. arborescens*.

Family / Class	Important Value Index			
	<i>M. gigantea</i>	<i>M. pruinosa</i>	<i>C. arborescens</i>	<i>A. crassicarpa</i>
Formicidae 1	32.8	39.68	6.43	52.94
Formicidae 2	29.37	31.01	3.81	55.71
Formicidae 3		9.61	10.37	
Formicidae 4	30.28	44.24	39.91	
Formicidae 5			30.98	
Curculionidae	24.04	16.1	11.55	5.04
Carabidae 1	17.8		8.93	5.4
Carabidae 2				5.4
Blatidae1	17.8	36.31	30.98	22.38
Blatidae 2	23.14		24.42	
Blatidae 3			6.43	
Polydesmidae	11.57	16.35		25.08
Rhinotermidae	29.37	21.15		77.22
Arachnida 1		4.81	11.43	
Arachnida 2		9.61		
Gryllidae		6.49		
Sapygidae			6.43	
Theriidae				21.98
Melolonthidae				14.21
Coicidae				16.59
Collembola	6.24	12.98	18.11	29.37

The second dominant group was Rhinotermidae and only be found in *A. crassicarpa*, *M. pruinosa*, and *M. gigantea*. This result was supported study that mentioned that the number of Rhinotermidae increases in peat land but decreases in mineral land and in contrast, the number of Termitidae enhance in mineral land but decline in peat land [40,41]. Together with Formicidae, this termite makes tunnels within the ground which increase the porosity and room for plant's root to penetrate the soil so that these two groups could be used as bioindicator. Another factor that leads the abundance of Formicidae is the availability organic materials, such as wood. Wood is the main resources for Rhinotermidae for food and shelters [37]. In addition, the highest number of Rhinotermidae found at *A. crassicarpa* supported a study conducted by Haneda et. al. [40] and Kirton et. al. [42] that showed termites were the main pests in *A. crassicarpa* and *A. mangium*. Yet as it turns out, no Rhinotermidae collected at *C. arborescens* presumably caused by lack of resources could be gathered under the stands. Moreover, a study conducted by Daviyana et. al. [43] stated that the extracts secreted from *C. arborescens* contain substances that control termites. In other words, inappropriate condition, which is harmful for termite, becomes limited factors for termite to live under *C. arborescens*.

In addition, it turns out that there was significantly different between diversity index value exotic tree species (*A. crassicarpa*) to other three native trees species ($p > 0.05$) (Table 5). Those native trees species had diversity index more than 2 and it means that the diversity level of those three native tree species was categorized as medium. In contrast, *A. crassicarpa* had the lowest diversity index value which valued 0.57 and was categorized as low. One thing that is interesting was the number of subclass collembola which was dominant at *A. crassicarpa*. Many studies reported that under dense canopy, high humidity, and thick litter, population of collembola was highly found [6,44,45]. In contrast, the population decline in open area which is heavily exposed to light [44,46]. Hence, it could be inferred that soil condition, such as humidity and litter thickness, formed by *A. crassicarpa* was more suitable for collembola to live compare to other native trees, particularly *M. gigantea*. Those studies also supported the result which found that the

lowest number of collembola was in *M. gigantea*. Based on observation, condition of *M. gigantea*'s floor was relatively clear from litter and was heavily exposed to sun light penetration compare to *A. crassicarpa*.

Table 5. Diversity index of understory's vegetation of *A. crassicarpa*, *M. gigantea*, *M. pruinosa*, and *C. arborescens*.

Species	Diversity index (shanon-weiner)
<i>M. gigantea</i>	2.12 ± 0.01a
<i>M. pruinosa</i>	2.22 ± 0.18a
<i>C. arborescens</i>	2.13 ± 0.18a
<i>A. crassicarpa</i>	0.57 ± 0.11b

Remark: the alphabets appear after the value showed a significant value ($p < 0.05$)

Analysis of variance of macrofauna's diversity showed that diversity index level at *A. crassicarpa* was significantly low than any other three native trees species ($p < 0.05$). Moreover, the diversity index was categorized low at *A. crassicarpa*. Meanwhile, diversity index was categorized as moderate at other three native trees species. This result was different to a study conducted by Handayani and Winara [37] that showed that diversity index at secondary forest of *A. crassicarpa* was at moderate level ($H' = 2.09$) and the lowest was at oil palm plantation ($H' = 0.73$). The value of diversity index at *A. crassicarpa* was similar to after-burned peat land di Riau [47]. Nevertheless, this result showed that there is transition and changing of macrofauna diversity level that could be seen from the differences of macrofauna's diversity level at three native trees species and exotic tree species.

4 Conclusions

Neprolephis biserrata and *Stenochlaenapalustris* dominated the understory vegetation in *A. crassicarpa*, *M. gigantea*, *M. pruinosa*, and *C. arborescens*. Meanwhile, narrow and broad leaves group dominantly varied between *A. crassicarpa*, *M. gigantea*, *M. pruinosa*, and *C. arborescens*. The diversity index in vegetation structure among those four tree species was not significant. Yet, *A. crassicarpa* had the lower value of diversity index. Moreover, the percentage of understory coverage under *A. crassicarpa* was significantly higher than that under all native tree species. On the other hand, formicidae and rhinotermitidae were dominant in *M. gigantea* and *A. crassicarpa*. Meanwhile, formicidae and blattidae were high in *M. pruinosa* and *C. arborescens*. Furthermore, diversity index of macrofauna were significantly low at *A. crassicarpa*'s compare to other three native trees species. This study informed that the introduction of *A. crassicarpa* give significant effect on the level of the understory's macrofauna biodiversity.

References

1. Von Rintelen K, Arida E, and Häuser C. A review of biodiversity-related issues and challenges in megadiverse Indonesia and other Southeast Asian countries. *Res Ideas Outcomes*. 2017 Sep;3:e20860.
2. Convention on Biological Diversity. Biodiversity facts; status and trends of biodiversity, including benefits from biodiversity and ecosystem services. Indonesia - Main Details. 2020.
3. Dirjen KSDAE. Statistik Dirjen KSDAE. Jakarta: Kementerian Lingkungan Hidup dan Kehutanan; 2017.
4. Wahyunto, Ritung S, Suparto, and Subagio H. Peatland distribution and its content in Sumatra and Kalimantan. Bogor; 2005.
5. Agus F, June T, Komara H, Syahbuddin H, Runtunuwu E, dan Susanti E. Mitigasi dan adaptasi perubahan iklim dari lahan perkebunan. Bogor; 2008.
6. Lisnawati Y, Suprijo H, dan Poedjirahajoe E. Hubungan kedekatan ekologis antara fauna tanah dengan karakteristik tanah gambut yang didrainase untuk HTI *Acacia crassicarpa*. *J. Manusia dan Lingkungan*. 2014;21(2):170–8.
7. Ariyanto DP, Sumarno, Supriyono, Yunus A, Pudjiasmanto B, and Rahayu. The productivity increasing of peatlands on community land by multi-cropping model in Riau Indonesia. *IOP Conf Ser Earth Environ Sci*. 2019;393(1).
8. Hasanah A and Setiawan MF. Rewetting design for tropical peatland restoration. *Sociae Polites*. 2020;21(2):111–25.

9. Hergoualc'h K, Atmadja S, Carmenta R, Martius C, Murdiyarso D, and Purnomo H. Managing peatlands in Indonesia: Challenges and opportunities for local and global communities. 2017.
10. Chin SY and Parish F. Peatlands: Status, challenges and actions in Southeast Asia. ASEAN Biodiversity. 2013 Jan;10.
11. Austin KG, Schwantes A, Gu Y, and Kasibhatla PS. What causes deforestation in Indonesia? Environ Res Lett. 2019;14(2). Sunardi S, Sulistijorini S, and Setyawati T. Invasion of *Acacia decurrens* willd after eruption of Mount Merapi, Indonesia. Biotropia (Bogor). 2017 Jan;24:35–46.
12. Sunardi S, Sulistijorini S, and Setyawati T. Invasion of *Acacia decurrens* willd after eruption of Mount Merapi, Indonesia. Biotropia (Bogor). 2017 Jan;24:35–46.
13. Montesinos D, Castro S, and Rodríguez-Echeverría S. Two invasive acacia species secure generalist pollinators in invaded communities. Acta Oecologica. 2016 Jul;74:In press.
14. Sutedjo S dan Warsudi W. Menakar sifat invasif spesies akasia mangium (*Acacia mangium* willd.) di hutan penelitian dan pendidikan bukit soeharto. J Hutan Trop. 2017;1(1):82–9.
15. Djufri. Pengaruh tegakan akasia (*Acacia nilotica*) (L.) willd. ex. del. terhadap komposisi dan keanekaragaman tumbuhan bawah di savana Balanan Taman Nasional Baluran Jawa Timur. J Biol Edukasi. 2011;3(2):38–50.
16. Doran C.J. Australian trees and shrubs: species for land rehabilitation and farm planting in the tropics. Canberra; 1997.
17. Nikmah N, Wiryani E. Struktur komposisi tumbuhan bawah tegakan jati di kebun benih klon (kbc) Padangan Bojonegoro. J Biol. 2016;5(1):30–8.
18. Indriyani L, Flamin A, dan Erna. Analisis keanekaragaman jenis tumbuhan bawah di hutan lindung jompi (Kelurahan Wali Kecamatan Watopute Kabupaten Muna Sulawesi Tenggara). Ecogreen. 2017;3(1):49–58.
19. Nahmani J, Lavelle P, dan Rossi JP. Does changing the taxonomical resolution alter the value of soil macro invertebrate as bioindicators of metal pollution? Soil Biol Biochem. 2006;38:385–396.
20. Junaedi A. Growth performance of three native tree species for pulpwood plantation in drained peatland of Pelalawan District, Riau. Indones J For Res. 2018;5(2):119–32.
21. Husna H, Wigena IGP, Dariah A, Marwanto S, Setyanto P, and Agus F. CO₂ emissions from tropical drained peat in Sumatra, Indonesia. Mitig Adapt Strateg Glob Chang. 2014;19(6):845–62.
22. Pribadi A dan Anggraeni I. Jenis dan struktur gulma pada tegakan *Acacia crassicaarpa* di lahan gambut. Tekno Hutan Tanam. 2010;4(1):33–40.
23. Supangat A., Pribadi A, Kosasih, dan Simatupang AD. Pengaruh pembangunan hutan tanaman terhadap biodiversitas. Laporan Hasil Penelitian BPHPS Kuok. Bangkinang; 2009.
24. Thomy Z, Masykur, and Yasmin Y. Diversity of plant species in Tripa Peat Swamp Forest, Aceh. In: Abs Sem Nas Masy Biodiv Indonesia. 2016. p. 89–131.
25. Giesen W and Sari ENN. Tropical peatland restoration report : the Indonesian case tropical peatland restoration report : The Indonesian Case Berbak Green Prosperity Partnership/Kemitraan Kesejahteraan Hijau (Kehijau Berbak). 2018;(3):99.
26. Daramola OS, Adeyemi OR, Adigun JA, and Adejuyigbe CO. Influence of row spacing and weed control methods on weed population dynamics in soybean (*Glycine max* L.). Int J Pest Manag. 2020;0(0):1–16.
27. Mirmanto E. Vegetation analyses of Sebangau peat swamp forest, Central Kalimantan. Biodiversitas J Biol Divers. 2009;11(2):82–8.
28. International C. Invasive Species Compendium. *Acacia crassicaarpa* (northern wattle). 2021.
29. Jambul R, Limin A, Ali AN, and Slik F. Invasive *Acacia mangium* dominance as an indicator for heath forest disturbance. Environ Sustain Indic. 2020;8(June):0–7.
30. Kirno F, Astiani D, dan Ekamawanti HA. Keanekaragaman Jenis tumbuhan paku-pakuan (pteridophyta) dan kondisi tempat tumbuhnya pada hutan rawa gambut sekunder dan lahan gambut terbuka. J Hutan Lestari. 2019;7(1):11–20.
31. Betty J, Linda R, dan Lovadi I. Inventarisasi jenis paku-pakuan (Pteridophyta) terrestrial di hutan dusun Tauk kecamatan Air Besar kabupaten Landak. Protobiont. 2015;4(1):94–102.
32. Tjitrosoepomo G. Taksonomi Tumbuhan : Schizophyta, Thallophyta, Bryophyta, Pteridophyta. Yogyakarta: Gajah Mada University Press;
33. Krebs CJ. Ecological Methodology. New York: Harper Collins Publisher; 1989.
34. Bela M, Ayu TMO. I, dan Vera M. Regenerasi hutan gambut pada kawasan lahan gambut bekas terbakar di Desa Pasir dan Desa Sungai Pinyuh, Kabupaten Mempawah, Kalimantan Barat. Pontianak; 2019.
35. Jackson DR and Finley J. Controlling understory fern competition for regeneration success. College of Agricultural Sciences, The Pennsylvania State University. 2016.
36. Yii JE, Bong CF, King JHP, and Jugah, K. Feeding preferences of oil palm pest subterranean

- termite *Coptotermes curvignathus* (Isoptera: Rhinotermitidae). *J. of Ento.* 2016; 13. 1-10
37. Handayani W and Winara, A. Keanekaragaman makrofauna tanah pada beberapa penggunaan lahan gambut. *J. Agroforestri Indonesia.* 2020; 3(2). 77 – 88
 38. Dewi FK, Djajakirana G, dan Suwardi. Kelimpahan dan keanekaragaman makrofauna tanah pada tiga penggunaan lahan gambut di Teluk Meranti, Kabupaten Pelalawan, Provinsi Riau. IPB University; 2016.
 39. Boror DJ, Triplehorn CA, dan Johnson NF. Pengenalan pelajaran serangga; diterjemahkan oleh S. Partossoedjono. Edisi ke-6. Yogyakarta: UGM Press; 1992.
 40. Haneda NF, Retmadhona IY, Nandika D, and Arinana. Biodiversity of subterranean termites on the *Acacia crassiparpa* plantation. *Biodiversitas.* 2017;18(4):1657–62.
 41. Cheng S, Kirton L., and Gurmit S. Termite attack on oil palm grown on peat soil: identification of pest status and factor contributing to the problem. *Planter.* 2008;84:200-210.
 42. Kirton LG, Brown VK, and Azmi M. The pest status of the termite *Coptotermes curvignathus* in *Acacia mangium* plantations: incidence, mode of attack and inherent predisposing factors. *J Trop For Sci.* 1999;11(4):822–31.
 43. Daviyana SA, Wardenaar E, and Yanti H. Pemanfaatan ekstrak kulit kayu gerunggang (*Cratoxylon arborescens*) untuk pengawetan kayu karet (*Hevea brasiliensis*) dari serangan rayap tanah. *J Hutan Lestari.* 2013;1(2):199–207.
 44. Widyastuti R, Santosa DA, dan Djajakirana G. The Diversity and abundance of springtail (collembola) on forests and smallholder in Jambi. *J Trop Soils.* 2015;20(3):173–80.
 45. Yan X, Ni Z, Chang L, Wang K, and Wu D. Soil warming elevates the abundance of collembola in the Songnen plain of China. *Sustain.* 2015;7(2):1161–71.
 46. Verhoef H and Selm A. Distribution and population dynamics of collembola in relation to soil moisture. *Ecography (Cop).* 2006 Jun;6:387–8.
 47. Gesriantuti N, Trantiati R, dan Badrun Y. Keanekaragaman serangga permukaan tanah pada lahan gambut bekas kebakaran dan Hutan Lindung Rokan Hulu, Provinsi Riau. *J. Phot.* 2016;7(1):147–155.