# An Overview- Advances in Chromatographic Techniques in Phytochemistry

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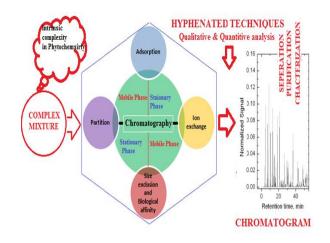
Abstract. The basic idea of the Chromatographic process is that the distinct compounds have different properties like solubility, adsorption, ion exchange, and affinity that can be regulated through various separation mechanisms causing the analytes to travel through the stationary phase at different speeds, resulting in their separation from a complex mixture. The Chromatographic techniques have been explored as multidimensional, which has been growing and evolving due to continuous advancements in technology allowing it to meet the upcoming challenges and analytical demands in various scientific scenarios including academics and industries. Phytochemistry is one of the oldest core areas of research in science where one of the major challenges in drug discovery from natural sources is the identification and isolation of closely related active molecules within these complex to the observed biological activities therefore there is utmost need to better understand their intrinsic complexity and exploit their vast commercial potential. Nowadays the hyphenated techniques with one component as the chromatographic technique have been widely explored as inevitable analytical tools for early detection and identification of bioactive compounds and driving forces in the evolution and discoveries in phytochemistry from crude plant extracts that boosted mainly due to rapid technological advancements in instrumentation. It's also significant as an intellectual method for phytotherapeutic quality control and regulation. In the present review, we have discussed the advances and insight of chromatographic techniques which can be explored for comprehensive chemical profiles of herbal medicine preparations or extracts in phytochemistry. The review provides a systematic update of the recent advancement and published approaches to methodology in phytochemistry.

**Keywords:** Gas chromatography, High-performance liquid chromatography, High-performance thin layer chromatography, supercritical fluid chromatography, Phytochemistry.

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#### **Graphical abstract:**



### 1. Introduction:

Chromatography is the most widely used separation technique in laboratories, where it can be explored for analysis, isolation, and purification. Chromatography has evolved from a simple technique for separating pigments into complex procedures involved in resolving the most difficult analytical and purifying challenges in separation science like phytochemistry during the last century [1]. The basic idea of the chromatographic process is that the distinct compounds have different properties like absorption, solubility, ion exchange, and affinity that can be regulated and explored through various separation mechanisms causing analytes to travel through the stationary phase at different speeds, resulting in their separation from a complex mixture [2]. Therefore the basic mechanism of any chromatographic procedure is to optimize standardized conditions where all the analytes of interest in the complex sample mixture move through the chromatographic system, but at differential speeds from the analytical column during elution to have sufficient separation, detection, and quantification from one another of the respective mixture. Indeed to enact the chromatographic process, the primary crucial function of the stationary phase is to retain or arrest analyte movement and that of the mobile phase is to force and hold the analyte to move from through the chromatographic system to the exit of the column [3]. It has been commonly used in the chemical manufacturing process industry in small and large-scale production. Chromatography has several significant applications including in the pharmaceutical sectors, food industry, molecular biology, biotechnology, and also in chemical industries due to its versatility in operating graphic techniques coupled with a reasonably well-developed framework under various variants and simplicity of approach [4].

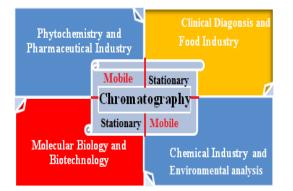


Fig. 1 Application spectrum of Chromatographic Techniques[5]

## 2. Current Condition of Ecological Phytochemistry:

Two main challenging factors that affect the phytochemical studies are the quantity of compound confined and its cleanliness. Both of these aspects have a significant impact on the possibility of composition analysis and the usage of molecule in vitro assays - something that needs to be considered. In recent decades, it has become increasingly significant. From its inception to the present, chromatography has evolved similarly to spectrophotometry. Furthermore, the difference between early analytical instruments like Thin Layer Chromatography (TLC) on paper and silica gel and current chromatographic technology is several orders of magnitude. Advances in chromatography such as ion exchange, affinity, exclusion, chiral, GC, HPLC, and counter-current (CCC) have expanded the options and reduced the amount of substance needed for analysis (e.g., Schreier et al., 1998; Hostettmann et al., 1997)[6]. Because biosynthetic routes could not be established without knowledge of the structures of small naturally occurring substances, the procedures mentioned above are particularly important. Without procedures that allow for the isolation of significant amounts of chemicals with a high degree of purity to run the corresponding biosensors, the research on the role of secondary metabolites could not be fully addressed. In this regard, developments in experimental and computational techniques have significantly impacted the focus of phytochemical ecosystems from a qualitative methodology to a wide perspective of their role inside the plant or concerning (biotic and abiotic) environmental elements [7].

## 3. Evolution and advances in Chromatography

Column adsorption chromatography was introduced by Mikhail Tswett at the beginning of the twentieth century, initially for the separation of plant pigments. Thereafter continue advancements in technology and instrumentation in the 20th century allowed chromatography to meet the upcoming challenges and analytical demands in a range of scientific scenarios including academics and industries [8]. Significant advances in chromatographic techniques as per timeline have been listed in the table 1The history of chromatography may be alienated into 3 key steps: the original starter of chromatography, In the 1950s, Martin's innovation, and in the 1970s, the primer of high-performance liquid chromatography systems [9].

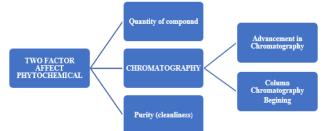


Fig. 2 Major challenges in phytochemistry and its influence on the evolution of chromatography[10]

Chromatographic Technique	Mechanism Separation	Time line	Inventor
Column Chromatography	Adsorption.	1906	Mikhail
		1000	Tswett
Thin-layer	Adsorption, Partition	1938	Izmailov and Shraiber
Chromatography(TLC) Paper chromatography	Adsorption, partition	1943	Martin and
Taper enromatography	Ausorption, partition	1945	Synge
Gas Liquid Chromatography	Adsorption, Partition.	1950	A.T. James
(GC)			and A.J.P. Martin
Size exclusion chromatography	Size and dimension of analyte	1955	Grant Henry
(SEC)			Lathe and
			Colin R Ruthven
Ion exchange chromatography	Ion exchange	1956	Sir Thompson
(IEC)	Ton exchange	1750	and J T Way
Affinity chromatography	Biological Affinity	1968	Cuatrecasas,
			Wilchek and
		1070	Anfinsen
High-performance liquid chromatography (HPLC)	Adsorption, Partition	1970	Csaba Horváth
Hydrophobic interaction	Adsorption, partition	1973	Hjerten
chromatography (HIC)		1070	G 1 G1 111
Ion pair chromatography	Ion pair formation, Ion interaction	1973	Gordon Shill
Capillary electrochromatography (CEC)	Follow both mechanisms of high- performance liquid	1974	Pretorius
electrochromatography (CEC)	chromatography (HPLC)		
Supercritical fluid	Adsorption, Partition	1980	Klesper
chromatography (SFC)	<b>r r r r</b>		· · · <b>r</b> ·
Ultra-performance liquid	Adsorption, Partition	2004	Waters
chromatography (UPLC)			

 Table 1. The advances in chromatographic techniques as per the time line[11][12][13]

Phytochemistry is one of the oldest core areas of research in science where one of the major challenges is the identification, isolation, and characterization of closely related active molecules within these complex matrices to the observed biological activities and drug discovery from natural sources, therefore there is a dire need to better understand their intrinsic complexity and exploit their vast commercial potential of natural products and their plant extracts [14]. Nowadays the hyphenated techniques with one of the components as a

chromatographic technique have been widely explored as an inevitable analytical tool for early detection and identification of bioactive compounds and driving forces in the evolution and drug discoveries in phytochemistry from crude plant extracts that are boosted mainly due rapid technological advancements in instrumentation [15]. Hyphenated techniques have evolved as a tool to find complete biological shapes of herbal medicine arrangements or quotations where the coupling of different two or more analytical techniques to solve more complex analytical problems where at least one of them is a separation technique while the other one is a spectroscopic detection technique. The goal of the coupling is to extract the material of a complex mixture for identification and quantification compared to that with only a logical method. Separation techniques include (GC) gas chromatography, (LC) liquid chromatography, (HPLC) high-performance liquid chromatography, and (CE) capillary electrophoresis [16]. In various traditional medicine systems like Ayurveda and traditional Chinese the medicinal properties are due to the occurrence of numerous kinds of bio-vigorous species and their disparity moreover qualitative or quantitative heavily impact the compressive substance outline of the herb-like loss and decrease in potency, medicinal properties, and increased toxicity. Therefore advances in chromatography provide ample technique of optimal for the excellent switch of greatest of the earliest and other explored herbal medicinal plant and their extract from TLC or paper chromatography to online detection methodology of bioactive molecules [17].

Type of	Hyphenated	Coupling with a suitable interface		Advantages	
Hyphenated Techniques	Techniques	Chromatographic technique	Spectroscopic technique		
Double hyphenated techniques.	LC -MS	LC	MS		
	LC-NMR	LC	NMR		
	LC-IR	LC	IR	Analysis:	
	CEMS	CE	MS	Accurate and Fast	
	GCIR	GC	IR		
	GC-MS	GC	MS	Automation:	
	GC-FTIR	GC	FTIR	Higher degree	
	HPLC-DAD	HPLC	MS		
Triple hyphenated techniques.	LC-UV-NMR-MS-	LC	UV-NMR	Sample	
	ESI		MS	throughput:	
	LC-API-MS	LC	MS	Higher	
	LC-ESI-MS	LC	MS	Reproducibility:	
	LC-NMR-MS	LC	NMR	Better	
	LC-DAD-API-MS	LC	MS NMR MS	Contamination: Reduce due to its closed system.	
	LC-PDA-MS	LC			
	LC-PDA-NMR-MS	LC	INIVIA IVIS		
				Quantification and	
				separation at the	
				same time.	

 Table 2. Hyphenated technique: Coupling of some chromatographic techniques and spectroscopic method with suitable interface[18][19][20][21][22]

Hyphenated techniques like HPLC coupled to mass and (LC-UV-MS) UV spectrometry are tremendously informative for a rapid survey of biological screening of natural products and plant extract. Nowadays, the two most commonly explored interfaces for usual creation investigation, are electrospray ionization (ESI) and atmospheric pressure chemical ionization (APCI). It is hard to cover all components of chromatography in a study of this scope, so a few key areas will be discussed in this review paper, such as Column, Ion exchange, Gel permeation, Affinity, HPTLC, and HPLC [23]. Combination approaches are the most recent advances in the evaluation of phytochemical compounds present in various trace levels (coupled techniques). Separation strategies are coupled with various detection systems in

these techniques. Chromatographic methods (e.g., GC, LC) is mostly used for separation, while spectroscopic methods (e.g., NMR, MS) is primarily used for detection [24]. Combinations of different chromatographic processes are also included in combined procedures. The combined technique should be discriminating for the intermediates to be determined, responsive over a broad concentration range, and allow for the most primary sources of the chemicals to be determined[25]. The origin of the analyzer, the simplicity with which the various methods can be combined, the sensitivity of the required results, and the availability of the equipment should all factor into the selection of the best combination methodology. The flavonoids and phenolic acids present in Flos Lamii albi were analyzed using high-performance liquid chromatography and high-performance thin-layer chromatography [26].

## 4. Applications of Chromatography in Phytochemistry

4.1. High-Performance Thin Layer Chromatography (HPTLC)

HPTLC is an enhanced form of thin-layer chromatography and is required to apply due to several TLC drawbacks, including occasional repeatability issues, and a lack of quantification precision. HPLC and GC, on the other hand, are discriminating, and not all of the chemicals in the sample are shown. Stahl's work, which established the consumption of gypsum as a stabilizing agent, chromatographic development, and normalized coating wideness, was a huge advancement in TLC (Stahl, 1958) [27]. The approach TLC provides aesthetic effects, but it is also simple and inexpensive. Samples may be analyzed in parallel, sample capability is great, and conclusions are produced quickly. TLC is adaptable, allowing for various identification. It's a suitable screening approach for biotic and organic inquiry, provided that credentials and partial results, as well as quantitative and semi-quantitative determinations of possible contamination. TLC bioautography can be used to test for biologically active compounds in association with microbes and other biological agents. TLC has several drawbacks, including occasional repeatability issues, and a lack of quantification precision [28].

TLC, on the other hand, will continue to be a quick and easy micro chromatography process. In HPTLC, the plates are attached to an inert phase with a scope of particles 5µm. The saucers provide superior phase separation and repeatability than standard TLC plates (size particles is 12µm), and also give more sensitive detection. There is a need for shorter development distances. The hypothetical number of plates is in the 5000 range, whereas the HPLC range is 6-10,000. HPTLC has a lesser separation power than HPLC, therefore later it is recommended for quantitative analysis [29]. Merck also sells HPTLC cups with subdivisions having spherical nature, which allows for quicker chromatography and greater separation. Merck offers water-resistant, plating and these plates may be utilized with 100 percent water for RP-18 W supports. HPTLC is also suitable for screening plant extracts before HPLC analysis [30]. Steroids, terpenoids, flavonoids, alkaloids, glycosides, sugar, and alkaloids were found in methanolic extracts of A. lanata Linn. According to preliminary phytochemical analysis. Carbohydrates, proteins, ash levels, and amino acids are all factors to consider. Different mobile phase compositions for HPTLC analyses were put to the test to achieve great resolution and precision. Peaks that are repeatable [31]. The intended result was attained by utilizing the mobile phase n-hexane-ethyl acetate (7.2:2.8). A. lanata contained 27 different forms of terpenoids, each with a different Rf value ranging from 0.06 to 0.97. Eight of the eleven different terpenoids found in leaves are solely found in leaves. In the root and

reproductive portions, eight different kinds of terpenoids were discovered (flowers-seeds) [32]. Five of the eight terpenoids found in the underground part of the root are exclusive to the root and are not found in the plant's aerial parts. Only the reproductive portions contain two distinct terpenoids with Rf values of 0.53, and 0.64. Terpenoids with Rf values of 0.69 and 0.76, for example, are only found in the stem. The terpenoid with the Rf value of 0.41 is found in abundance in all of the plant's aerial parts (stem, leaves, and reproductive parts) [33].

#### 4.2. High-Performance Liquid Chromatography

Even with the fact that HPLC has only been around for nearly 40 years, the technology has seen some of the most significant advancements in chromatography. The time 1967 was a watershed moment in the starter of HPLC, through articles by Scott, Horvath, and Huber, however, Moore and co-workers described the first automated liquid chromatograph with descent absorbance in 1958[34]. Supreme phytonutrient departures were done using thinlayer and open-layer chromatography before HPLC. Open-column was tedious, costly, laborintensive, and needed a lot of tasting. Very tiny samples could be analyzed with paper chromatography and TLC, and the precision and consistency were enhanced [35]. Quantitation, on the other hand, was still lacking, and the purpose of comparable molecules was challenging. Although gas chromatography gave good resolution, it was limited to volatile materials (only about carbon-based chemicals 20% can be alienated by vapor chromatography), necessitating recognition [36]. Liquid-fathomable, roasting versatile, nonadaptable chemicals have to be separated quickly, and with spatial precision. With the capacity to segregate, detect, and analyze the chemicals contained in any trial which can be disbanded in a liquescent, HPLC has become one of the most successful methods in analytical chemistry. The thickness of liquids is 100 times that of gases, requiring the use of heaviness in the columns and giving rise to the term "high-pressure liquid chromatography." As the constituent part became less significant and pillars suited squatter, "pressure" was displaced by "efficiency." The huge range of motionless and moveable stages should provide you with a lot of flexibility in terms of determining the best separation circumstances [37]. However, only rather big particles were initially accessible. The primer of tiny permeable subdivisions named silica with a span of about 10µm, as well as the manufacture of chemically bound phases, such as octyl (RP8) materials and reversed-phase (RP) octadecyl (RP18) completely transformed the situation. Stabilization of silica surfaces by specified alkylation, use of ultrapure silica, and advancements in bonding and side techniques have resulted in very adaptable reversed-phase systems with precision, isolation power, durability, reliability, and efficiency. By far the most extensively utilized chromatographic technology is highperformance liquid chromatography [38]. HPLC is divided into numerous categories, including flipping, thermal decomposition, and immobilized enzyme reactor. HPLC can take several forms, both in terms of what it does and how it is carried out. This has a significant impact on how and when HPLC procedures are used, however, HPLC has proved to be beneficial in both clinical and medicinal applications [39]. Adrenaline and dopamine are Catecholamines that are significant for a diversity of natural processes. For example, Diseases containing Parkinson's disease, heart-related infection, and genetic disease can be recognized by examining their ancestor and metabolites like biotransformation. Adrenaline and dopamine are catecholamines that are required for a range of biological processes [40]. The antecedents and metabolites of diseases comprising paralysis agitans, heart-related problems, and genetic problems can be used to analyze them. HPLC is the best way to isolate and analyze particles than other different methods, making it a tremendous challenge for such therapeutic and indicative presentations. Measurement of confinement time is used to categorize compounds in HPLC. It takes time for the molecule to permit over and done by a

column lined with adsorbents that cooperate in an altered way with different types of molecules is considered as reservation time. This ended in an assortment of circumstances. The prospective use of RP-HPLC in diagnostic frameworks was unproven in 1976[41]. Researchers used hydrophobic characteristics to speed up the procedure by separating catecholamine metabolites and amines in the same way. This is a delicate balance to a pH interaction since acidic catecholamine metabolites are maintained for prolonged at lower pH values, whilst amines are held for longer at higher pH levels. A cancer name Pheochromocytoma is interconnected with the compassionate anxious scheme that can cause death. It is generated from neural peak material, entailing that catecholamines are concealed. It can produce hypertension, which can make identification extra complicated because of the set-up of, maybe the solitary change between it and hypertension [42]. As medicament, was supplementary and extensively bent, the guideline was legislated to assure applicable fabrication and transparency of drugs scattered. HPLC is one of the greatest narrowly used techniques for formative treatment clarity all around the world. HPLC has shown to be operative in the medicine-related sector since its inauguration. Steroids, alkaloids, and antibiotics all were scrutinized using HPLC [43]. HPLC isn't only used for exploring completed therapeutical goods. HPLC is generally used in engineering because it can isolate kinds of stuff. In medicine-related experiments, it can demonstrate enantiomer molecular transparency is a need, and HPLC is superlative for that. Phenylcarbamate and Polysaccharide benzoate derivatives are the most often utilized CSPs in pharmaceutical chemistry [44].

#### 4.3 Gas Chromatography

Our environment is made up of a variety of complicated mixes. Petroleum might include over 100,000 constituents. The human body is considered to have on the order of 100,000 distinct proteins [45]. Natural items, such as essential oils, are frequently complicated as well. Severance methods are required to analyze them, whereas even the simplest, but still tough, mixes seen in the pharmaceutical sector, for example, almost always necessitate chromatography or a comparable technique [46]. GC produces separations through a sequence of barriers between a stirring vapor period and a stagnant watery level kept in a compact aperture cylinder introduced after a mixture in a constricted ensemble. A gauge displays the alignment of the thrill torrent as it materializes from the separated component, and the consequential gestures serve as a data-logging interface [47]. Chemicals having boiling temperatures ranging from instantaneous 0k to 700K, or those being warmed to vapor stress of a scarce mmHg without breakdown, can be analyzed by GC. To boost fluctuation, this range is broadened through esterification. The model dimension can be as trivial as pg., but prefatory rather than methodical solicitations can be handled [48]. In numerous sectors, including petrochemical manufacturing, conservational, nourishment noxious waste, drug residue analysis, and pathological scrutiny, GC is a customary investigative technology that underlies exploration, enlargement, and superiority rheostat [49]. Gas chromatography (GC) is broadly used in the realm of sustenance exploration. The computable and/or quality-related examination of natural products, food conformation, food condiments, taste and fragrance mechanisms, and a range of other things are common uses pollutants, such as pesticide contaminants and transformation products, antiseptic, and pollutants in the environment physician medications, natural poisons, and packaging materials. Past and present trends in GC for food applications are evaluated, and future trends are forecasted [50]. Among the many novel methods being technologically advanced, the journalists anticipate that fast-GC/ (MS) mass spectrometry) will have the greatest influence on food analysis applications

during the next century [51]. (GC) Gas Chromatography transmits on to show an essential part in the documentation and assessment of environmental contaminants today. Techniques for sample preparation are given special consideration. Impulsive living compounds (ILCs), (PAHs) Polycyclic aromatic hydrocarbons, insecticides, and disinfectant chemicals are the different types of organic pollutants [52]. These last embrace polybrominated diphenyl ethers, polybrominated biphenyls, brominated flame retardants, terphenyls, polychlorinated biphenyls, naphthalenes and alkanes, organochlorine pesticides, dibenzofurans and polychlorinated dipenzopdioxins [53].

#### 4.4. Supercritical Fluid Chromatography

For investigative and refining solicitations, it provides an eco-friendly option. CO2 is employed as the polar solvent in SFC under exact pressure and temperature conditions. The critical components for hydrothermal Carbonization are 31C and 74bar [54]. CO2's physiochemical features give great perks: When compared to the solvents frequently employed in hplc analysis, CO2 is innocuous, fireproof, amicable, and has a lower budget. These characteristics are in line with biomass conversion standards, confirming the method's utility in the chemical analysis [55]. Expenditure of CO2 is precisely prevalent, because, after the spinal compression manager, the CO2 is not pressurized countenancing the assortment of strenuous segments without supplementary dynamism to eradicate the flush. It is known that SFC has developed one of its treasured performances for the assemblage of concerted portions of poisons such as Insect killer, 1, 1'-Biphenyl chloro derivatives, etc.[56].

Seeds K.uvaria were composed by Rechereche-LVMH. The mass of seeds is about 300gm. were everywhere with a Braun liquidizer of 1Lt. Seeds were mined by Supercritical Unsolidified abstraction with an SFE-500 gadget, with unalloyed CO2 at 333k and 290bar [57]. The withdrawal interval was an occupation of Figure of CO2 delivered in removal thriving. When the CO2 corpus grasped 3 kg, the pulling out was motionless. The emollient was sheltered and desiccated in emptiness with rough milliliters of liquor to expedite the ventilation [58].

	Chromatographic Techniques				
S.No		HPTLC	HPLC	GC	SFC
1	Plant	Fumaria parviflora	Leucas aspera	Thymus	Kniphofia
	species			vulgaris L	uvaria
2	Extracting	Combination of	Benzene,	Hexane and	Carbon dioxide
	Solvent	methanol, ethyl	Chloroform	isooctane	
	used	acetate, formic			
		acid, and toluene			
		(2:6:1:6)			
3	Detector	Fluorescent	UV- visible	Flame	Filled
	used	Detector	absorbance	ionization	Teflon./MS
			detector	sensor	
4	Lamp used	Mercury vapor,	Deuterium	PID	Mass
		Tungsten.			spectrometer.
5	Features	Exceedingly	High	Used to detach	Solute solubility
		effectual, little	compassion,	the chemical	increases with
		flush consumption	Small taster	amalgams of a	intensification
		per illustration	dimensions.	sample	in fluid density.
				mixture.	
6	Limitations	An inadequate	Cost,	Unsatisfactory	Low UV
		number of	involvedness	precariousness.	thoughtfulness.
		mockups per plate,			
		Little retreat bed.			

Table 3	<ul> <li>Application</li> </ul>	of chromatograph	y in phytochem	istry[59][60][61][62]

## Conclusion

Chromatography has evolved from a simple technique for separating pigments into a complex set of chromatographic procedures along with hyphenated techniques due to continuous technological advances that allow resolving the most difficult upcoming analytical and purifying challenges in scientific scenarios of academics and industries that provide one of the driving force in the evolution and drug discoveries in phytochemistry and beyond. Moreover, the hyphenated techniques have evolved as a tool to get inclusive biochemical outlines of herbal medicine provisions or excerpts where the coupling of different two (or more) analytical techniques to solve more complex analytical problems where at least one of them is a separation technique like chromatography while other one is spectroscopic detection technique. In hyphenated techniques, various separation techniques have been explored like capillary electrophoresis (CE), Liquid Chromatography (LC), Gas Chromatography (GC), and High-performance liquid Chromatograph (HPLC). Therefore advances in chromatography provide ample technique of selection for the excellent controller of the greatest antique and other explored herbal medicinal plant and their extract from TLC or paper chromatography to online chromatographic detection methodology for bioactive molecules. Without chromatography, phytochemistry would be a lot more difficult; analytical processes would be more complicated, and some issues, such as metabolomic analysis, would be impossible to solve. With multi-constituent mixtures of great complications to analyze, the problems of proteomics and metabolomics are already pervasive. Without a doubt, developments in HPLC via novel column technology, combined with mass spectrometry as an uncovering method, will give solutions to these difficulties. Characterization of the multifaceted plant mixes used in Chinese traditional medicine is also ongoing. This is required to meet the needs of regulatory agencies. There will be advancements in column packing as well. New chemistries will be created, and a higher level of stability will be sought.

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#### References

- [1] A. Marston, *Role of advances in chromatographic techniques in phytochemistry*, *Phytochemistry*, vol. 68, no. 22–24, (2007).doi: 10.1016/j.phytochem.2007.08.004
- [2] J. C. Bellot and J. S. Condoret, Modelling of liquid chromatography equilibria, Process Bioche., vol. 28, no. 6, pp. 365–376, (1993).doi: 10.1016/0032-9592(93)80023-A.
- [3] K. Ichi Kasai, *Trypsin and affinity chromatography, J. Chromatogr. A* vol. 597, no. 1–2, pp. 3–18, (1992).doi: 10.1016/0021-9673(92)80092-9.
- [4] D. Guillarme, D. T. T. Nguyen, S. Rudaz, and J. L. Veuthey, Method transfer for fast liquid chromatography in pharmaceutical analysis: Application to short columns packed with small particle. Part I: Isocratic separation, Eur. J. Pharm. Biopharm., vol. 66, no. 3, pp. 475–482, (2007). doi: 10.1016/j.ejpb.2006.11.027.
- [5] V. Mapelli, L. Olsson, and J. Nielsen, *Metabolic footprinting in microbiology: methods and applications in functional genomics and biotechnology, Trends Biotechnol.*, vol. 26, no. 9, pp. 490–497, (2008). doi: 10.1016/j.tibtech.2008.05.008.

- [6] E. González-Burgos, C. Fernández-Moriano, and M. P. Gómez-Serranillos, *Current knowledge on Parmelia genus: Ecological interest, phytochemistry, biological activities, and therapeutic potential, Phytochemistry,* vol. 165, no. June, p. 112051, (2019). doi: 10.1016/j.phytochem.2019.112051.
- [7] C. A. Lucy, *Recent advances in ion chromatography: A perspective, J. Chromatogr. A* vol. 739, no. 1–2, pp. 3–13, (1996). doi: 10.1016/0021-9673(96)00034-9.
- [8] F. A. Macías, J. L. G. Galindo, and J. C. G. Galindo, Evolution and current status of ecological phytochemistry, Phytochemistry, vol. 68, no. 22–24, pp. 2917–2936, (2007). doi: 10.1016/j.phytochem.2007.10.010.
- [9] J. C. Touchstone, *History of Chromatography, J. Liq. Chromatogr.*, vol. 16, no. 8, pp. 1647–1665, (1993). doi: 10.1080/10826079308021679.
- [10] J. Meija, M. Montes-Bayón, J. A. Caruso, and A. Sanz-Medel, Integrated mass spectrometry in (semi-)metal speciation and its potential in phytochemistry, TrAC -Trends Anal. Chem., vol. 25, no. 1, pp. 44–51, (2006).doi: 10.1016/j.trac.2005.04.003.
- M. Saito, *History of supercritical fluid chromatography: Instrumental development,* J. Biosci. Bioeng, vol. 115, no. 6, pp. 590–599, (2013). doi: 10.1016/j.jbiosc.2012.12.008.
- [12] E. Heftmann, *Survey of chromatography and electrophoresis*, *J. Chromatogr. Libr.*, vol. 22, no. PA, pp. A1–A18, (1983).doi: 10.1016/S0301-4770(08)60862-3.
- [13] R. O. B. Wijesekera and C. O. Chichester, The chemistry and technology of cinnamon, C R C Crit. Rev. Food Sci. Nutr., vol. 10, no. 1, pp. 1–30, (1978). doi: 10.1080/10408397809527243.
- [14] J. D. Phillipson, *Phytochemistry and pharmacognosy*, *Phytochemistry*, vol. 68, no. 22–24, pp. 2960–2972, (2007) doi: 10.1016/j.phytochem.2007.06.028.
- [15] G. Purcaro, S. Moret, and L. Conte, Hyphenated liquid chromatography-gas chromatography technique: Recent evolution and applications, J. Chromatogr. A, vol. 1255, pp. 100–111, (2012). doi: 10.1016/j.chroma.2012.02.018.
- [16] S. Gocan, Hyphenated techniques in thin-layer chromatography, *Adv. Chromatogr.*, vol. 47, pp. 353–444, (2009). doi: 10.1201/9781420060379.ch10.
- [17] G. Morlock and W. Schwack, Hyphenations in planar chromatography, J. Chromatogr. A vol. 1217, no. 43, pp. 6600–6609, (2010) doi: 10.1016/j.chroma.2010.04.058.
- [18] D. L. Norwood, J. O. Mullis, and T. N. Feinberg, 7 Hyphenated techniques, Sep. Sci. Technol., vol. 8, pp. 189–235, (2007). doi: 10.1016/S0149-6395(07)80013-4.
- [19] R. Lobiński and J. Szpunar, Biochemical speciation analysis by hyphenated techniques, Anal. Chim. Acta, vol. 400, no. 1–3, pp. 321–332, (1999). doi: 10.1016/S0003-2670(99)00628-5.
- [20] M. I. G. S. Almeida, J. M. Estela, and V. Cerdà, *Multisyringe flow injection potentialities for hyphenation with different types of separation techniques, Anal. Lett.*, vol. 44, no. 1–3, pp. 360–373, (2011). doi: 10.1080/00032719.2010.500779.
- [21] T. N. Feinberg, *Hyphenated characterization techniques, Sep. Sci. Technol.*, vol. 5, no. C, pp. 341–359, (2004). doi: 10.1016/S0149-6395(03)80015-5.
- [22] V. Räntzsch, M. Wilhelm, and G. Guthausen, *Hyphenated low-field NMR* techniques: Combining NMR with NIR, GPC/SEC and rheometry, Magn. Reson. Chem., vol. 54, no. 6, pp. 494–501, (2016). doi: 10.1002/mrc.4219.
- [23] H. Seifi, T. Gholami, S. Seifi, S. M. Ghoreishi, and M. Salavati-Niasari, A review on current trends in thermal analysis and hyphenated techniques in the investigation of physical, mechanical and chemical properties of nanomaterials, vol. 149. Elsevier B.V, (2020).

- [24] W. Lorenc, D. Kruszka, P. Kachlicki, J. Kozłowska, and D. Barałkiewicz, Arsenic species and their transformation pathways in marine plants. The usefulness of advanced hyphenated techniques HPLC/ICP-MS and UPLC/ESI-MS/MS in arsenic species analysis, Talanta, vol. 220, p. 121384, (2020). doi: 10.1016/j.talanta.2020.121384.
- [25] T. D.G., Novel Analytical Techniques used in Identification and Isolation of Impurities in Pharmaceuticals an Overview, J. Pharm. Sci. Res., vol. 12, no. 1, pp. 37–42, (2020).
- [26] W. Parys, M. Dołowy, and A. Pyka-Pająk, Current strategies for studying the natural and synthetic bioactive compounds in food by chromatographic separation techniques, *Processes*, vol. 9, no. 7, (2021). doi: 10.3390/pr9071100.
- [27] D. H. Shewiyo, E. Kaale, P. G. Risha, B. Dejaegher, J. Smeyers-Verbeke, and Y. Vander Heyden, *HPTLC methods to assay active ingredients in pharmaceutical formulations: A review of the method development and validation steps, J. Pharm. Biomed. Anal.*, vol. 66, pp. 11–23, (2012). doi: 10.1016/j.jpba.2012.03.034.
- [28] D. Kasote, A. Ahmad, W. Chen, S. Combrinck, and A. Viljoen, HPTLC-MS as an efficient hyphenated technique for the rapid identification of antimicrobial compounds from propolis, Phytochem. Lett., vol. 11, pp. 326–331, (2015). doi: 10.1016/j.phytol.2014.08.017.
- [29] P. Hariprasad and N. Ramakrishnan, *Chromatographic fingerprint analysis of Rumex vicarious L. by HPTLC technique, Asian Pac. J. Trop. Biomed.*, vol. 2, no. 1 SUPPL., pp. S57–S63, (2012). doi: 10.1016/S2221-1691(12)60130-0.
- [30] S. Sampathkumar and N. Ramakrishnan, Chromatographic fingerprint analysis of Naringi crenulata by HPTLC technique, Asian Pac. J. Trop. Biomed., vol. 1, no. SUPPL. 2, pp. S195–S198, (2011). doi: 10.1016/S2221-1691(11)60155-X.
- [31] Y. Mariswamy, W. E. Gnaraj, and J. M. Antonisamy, *Chromatographic fingerprint analysis on flavonoid constituents of the medicinally important plant Aerva lanata L. By HPTLC technique, Asian Pac. J. Trop. Biomed.*, vol. 1, no. SUPPL. 1, p. S8, (2011). doi: 10.1016/S2221-1691(11)60112-3.
- [32] S. Gupta and R. Gupta, Detection and quantification of quercetin in roots, leaves, and flowers of Clerodendrum infortunatum L., Asian Pacific J. Trop. Dis., vol. 2, no. SUPPL2, pp. S940–S943, (2012). doi: 10.1016/S2222-1808(12)60296-5.
- [33] C. Govindasamy and R. Srinivasan, In vitro antibacterial activity and phytochemical analysis of Catharanthus roseus (Linn.) G. Don., Asian Pac. J. Trop. Biomed., vol. 2, no. 1 SUPPL., pp. S155–S158, (2012). doi: 10.1016/S2221-1691(12)60148-8.
- [34] S. Fluoroquinolone, D. S. North, D. Pharm, D. N. Fish, D. Pharm, and J. J. Redington, *Evaluations of New Drugs*, (1998).
- [35] Anesth 1988 PK and Protein Binding with Propofol in Cirrhosis.pdf.".
- [36] K. N. Waliszewski, V. T. Pardio, and S. L. Ovando, A simple and rapid HPLC technique for vanillin determination in alcohol extract, Food Chem., vol. 101, no. 3, pp. 1059–1062, (2007). doi: 10.1016/j.foodchem.2006.03.004.
- [37] M. Marcinkowska and D. Barałkiewicz, Multielemental speciation analysis by advanced hyphenated technique – HPLC/ICP-MS: A review, *Talanta*, vol. 161, pp. 177–204, (2016). doi: 10.1016/j.talanta.2016.08.034.
- [38] G. K. J. Chao and J. C. Suatoni, Determination of phenolic compounds by HPLC, J. Chromatogr. Sci., vol. 20, no. 9, pp. 436–440, (1982). doi: 10.1093/chromsci/20.9.436.
- [39] D. Steinmann and M. Ganzera, Recent advances on HPLC/MS in medicinal plant analysis, J. Pharm. Biomed. Anal., vol. 55, no. 4, pp. 744–757, (2011). doi: 10.1016/j.jpba.2010.11.015.

- [40] G. Cimpan and S. Gocan, Analysis of medicinal plants by HPLC: Recent approaches, J. Liq. Chromatogr. Relat. Technol., vol. 25, no. 13–15, pp. 2225–2292, (2002). doi: 10.1081/JLC-120014003.
- [41] T. Yi, K. S. Y. Leung, G. H. Lu, H. Zhang, and K. Chan, Identification and determination of the major constituents in traditional Chinese medicinal plant polygonum multiflorum Thunb by HPLC coupled with PAD and ESI/MS, Phytochem. Anal., vol. 18, no. 3, pp. 181–187, (2007). doi: 10.1002/pca.963.
- [42] M. Vogeser and C. Seger, A decade of HPLC-MS/MS in the routine clinical laboratory - Goals for further developments, *Clin. Biochem.*, vol. 41, no. 9, pp. 649– 662, (2008). doi: 10.1016/j.clinbiochem.2008.02.017.
- [43] D. C. Costa, H. S. Costa, T. G. Albuquerque, F. Ramos, M. C. Castilho, and A. Sanches-Silva, Advances in phenolic compounds analysis of aromatic plants and their potential applications, Trends Food Sci. Technol., vol. 45, no. 2, pp. 336–354, (2015). doi: 10.1016/j.tifs.2015.06.009.
- [44] S. Görög, *The changing face of pharmaceutical analysis, TrAC Trends Anal. Chem.*, vol. 26, no. 1, pp. 12–17, (2007). doi: 10.1016/j.trac.2006.07.011.
- [45] H. Weil and T. I. Willams, *History of chromatography, Nature*, vol. 166, no. 4232, pp. 1000–1001, (1950) doi: 10.1038/1661000b0.
- [46] J. V. Seeley and S. K. Seeley, Multidimensional gas chromatography: Fundamental advances and new applications, Anal. Chem., vol. 85, no. 2, pp. 557–578, (2013) doi: 10.1021/ac303195u.
- [47] S. J. Lehotay and J. Hajšlová, Application of gas chromatography in food analysis, TrAC - Trends Anal. Chem., vol. 21, no. 9–10, pp. 686–697, (2002). doi: 10.1016/S0165-9936(02)00805-1.
- [48] F. J. Santos and M. T. Galceran, *The application of gas chromatography to environmental analysis, TrAC Trends Anal. Chem.*, vol. 21, no. 9–10, pp. 672–685, (2002). doi: 10.1016/S0165-9936(02)00813-0.
- [49] C. A. Cramers, H. G. Janssen, M. M. Van Deursen, and P. A. Leclercq, *High-speed gas chromatography: An overview of various concepts, J. Chromatogr. A* vol. 856, no. 1–2, pp. 315–329, (1999). doi: 10.1016/S0021-9673(99)00227-7.
- [50] P. Q. Tranchida, P. Donato, F. Cacciola, M. Beccaria, P. Dugo, and L. Mondello, *Potential of comprehensive chromatography in food analysis, TrAC - Trends Anal. Chem.*, vol. 52, pp. 186–205, (2013). doi: 10.1016/j.trac.2013.07.008.
- [51] M. Adahchour, J. Beens, R. J. J. Vreuls, A. M. Batenburg, and U. A. T. Brinkman, *Comprehensive two-dimensional gas chromatography of complex samples by using a 'reversed-type' column combination: Application to food analysis, J. Chromatogr. A* vol. 1054, no. 1–2, pp. 47–55, (2004) doi: 10.1016/j.chroma.2004.07.084.
- [52] P. M. Medeiros and B. R. T. Simoneit, *Analysis of sugars in environmental samples* by gas chromatography-mass spectrometry, J. Chromatogr. A, vol. 1141, no. 2, pp. 271–278, (2007) doi: 10.1016/j.chroma.2006.12.017.
- [53] T. Barri and J. Å. Jönsson, Advances and developments in membrane extraction for gas chromatography: Techniques and applications, J. Chromatogr. A vol. 1186, no. 1–2, pp. 16–38, (2008). doi: 10.1016/j.chroma.2008.02.002.
- [54] L. T. Taylor, *Supercritical fluid chromatography for the 21st century*, *J. Supercrit. Fluids*, vol. 47, no. 3, pp. 566–573, (2009) doi: 10.1016/j.supflu.2008.09.012.
- [55] V. Desfontaine, D. Guillarme, E. Francotte, and L. Nováková, Supercritical fluid chromatography in pharmaceutical analysis, J. Pharm. Biomed. Anal., vol. 113, pp. 56–71, (2015). doi: 10.1016/j.jpba.2015.03.007.
- [56] L. T. Taylor, *Supercritical fluid chromatography, Anal. Chem.*, vol. 82, no. 12, pp. 4925–4935, (2010) doi: 10.1021/ac101194x.

- [57] R. M. Hannan and H. H. Hill, Analysis of lipids in aging seed using capillary supercritical fluid chromatography, J. Chromatogr. A vol. 547, no. C, pp. 393–401, (1991) doi: 10.1016/S0021-9673(01)88662-3.
- [58] T. L. Chester, J. D. Pinkston, and D. E. Raynie, Supercritical fluid chromatography and extraction, Anal. Chem., vol. 68, no. 12, pp. 487–514, (1996). doi: 10.1021/a1960017i.
- [59] A. Marston, *Thin-layer chromatography with biological detection in phytochemistry, J. Chromatogr. A*, vol. 1218, no. 19, pp. 2676–2683, (2011). doi: 10.1016/j.chroma.2010.12.068.
- [60] Ł. M. Ciešla, M. Waksmundzka-Hajnos, K. A. Wojtunik, and M. Hajnos, *Thin-layer chromatography coupled with biological detection to screen natural mixtures for potential drug leads, Phytochem. Lett.*, vol. 11, pp. 445–454, (2015). doi: 10.1016/j.phytol.2015.02.005.
- [61] R. Gordillo, Supercritical fluid chromatography hyphenated to mass spectrometry for metabolomics applications, J. Sep. Sci., vol. 44, no. 1, pp. 448–463, (2021). doi: 10.1002/jssc.202000805.
- [62] L. Witte, P. Rubiolo, C. Bicchi, and T. Hartmann, Comparative analysis of pyrrolizidine alkaloids from natural sources by gas chromatography-mass spectrometry," Phytochemistry, vol. 32, no. 1, pp. 187–196, (1992). doi: 10.1016/0031-9422(92)80130-7.