# Characterization of Peppermint Plant Extract and Antimicrobial Activity

Donia Waleed Khaled1\*

<sup>1</sup>Department of Optical Techniques, AlNoor University College, Nineveh, Iraq

Abstract. The use of plant extracts in the medical and pharmacological fields have been increased in the last years. The plants have shown to possess materials with important features that can be used as a treatment in multiple health risks such as inflammation, cardiovascular disorders, bacteria and fungus. Peppermint was used extensively as medical herbs and have given a remarkable results. We aimed to prepare water and alcoholic (methanol) extract of peppermint leaves and characterize the presence of some phytochemical compounds qualitatively, along with some trace elements (copper, zinc and selenium). The antimicrobial activity of peppermint extract was investigated against two Gram negative escherichia coli as well as klebsiella sp, also two Gram positive bacterial strains Staphylococcus epidermidis as well as Staphylococcus aureus, as well as one fungi (Candida albicans). We have obtained the presence of flavonoids, polyphenolic compounds, steroids, carbohydrate, and protein contents in the extract of peppermint. Copper, zinc and selenium levels were detected in good amount in the extract. Furthermore, both water and methanol extract of peppermint have shown good antimicrobial activity against the experimented strains. Nevertheless, the extraction by methanol have shown more powerful activity than water against all strains.

Keywords: Peppermint, mentha piperita, antibacterial, phytochemicals.

## 1. Introduction

Plants have long been seen as a possible source of medicine, and have been utilized to treat a variety of maladies in primitive form. A range of bioactive chemicals found in various regions of plants has rekindled interest in producing an alternative medicine. The conventional medicinal herbs system has been utilized internationally since olden days, and as a result, a large body of literature on the antibacterial activity of many plant species is accessible [1].

<sup>\*</sup> Corresponding Author: researcherstaff08@alnoor.edu.iq

<sup>©</sup> The Authors, published by EDP Sciences. This is an open access article distributed under the terms of the Creative Commons Attribution License 4.0 (https://creativecommons.org/licenses/by/4.0/).

In the culinary, beverage, cosmetic, health, and tobacco industries, peppermint (Mentha Piperita) oil is frequently utilized [2-5]. Menthone, menthofuran, and menthol are the main components of peppermint oil. Peppermint oil has antioxidant effects [6, 7], antibacterial activity [8,] and is one of the most important ingredients in certain over-the-counter irritable bowel syndrome medicines in Europe [9]. Although both the extract and the leaves are listed as biological additions, only the extract is said to be employed. Peppermint Water is a flavoring ingredient or scent component, according to [10]. The volatile oil content of peppermint leaves ranges from 0.5 to 4%. Furthermore, polyphenolic components such as rosmarinic acid, flavone and flavanone glycosides, and both flavone and flavanone glycosides make up much to 21.7 percent of this extensively used herbal medication [11]. Peppermint oil has antibacterial efficacy against Aspergillus niger, Rhizopus solani, and Alternaria alternative [12], as well as Pseudomonas syringe, Xanthamonas campestris, E. coli, Pseudomonas aeruginosa, and Salmonella typhimurium [13]. Peppermint leaf juice had stronger antibacterial activity than peppermint stem juice against Gram negative bacteria (Saeed and Tariq, 2005). Peppermint oil had stronger antibacterial activity than S. aureus against Candida albicans and E. coli [14]. Because of the reports on the importance features of peppermint extract, we have designed this study to characterize the extract of peppermint leaf extract for phytochemicals identification, some trace elements presence such as copper, selenium, and zinc. Furthermore, we have investigated the antimicrobial effect of peppermint extract on four different types of microbes.

# 2. Materials and Methods

#### 2.1. Preparation of peppermint extract

Peppermint were purchased from the local market in Kut – Wasit – Iraq. The leaf of peppermint plants were washed to remove the dust and any other pollutants, then it was minced. A Weight of 1 g of the minced peppermint were added to 50mL of deionized water or methanol (Merk, Germany). The water or methanol mixtures of peppermint were shaked by using a waterbath with shaker under mild heat (50-55 °C) for 1h. Then, the beakers were allowed to stand at room temperature to cool down, then it was filtered, and the solution was stored for analyses.

## 2.2. Trace elements analyses

The determination of copper (Cu), Zinc (Zn), and Selenium (Se) were performed by using atomic absorption spectroscopy (AAS) approach. The analyses required a pre-step, in which 10mL of the water extract were mixed with 10 drops of HNO<sub>3</sub> (Merck, Germany) in a flash and covered with a glass sealer. The trace elements were measured by using Shimadzu AA-670, Flame AAS [15].

## 2.3. Phytochemical qualitative analyses

The chemical components of the produced fruit extract were detected using several assays according to the standard AOAC (1990) procedure [16]. Glycosides, alkaloids, saponins, phenolic substances, tannins, flavonoids, proteins, and steroids were among them.

## 2.4. Antimicrobial activity

Both methanol and water extracts of peppermint were examined against two Gram negative bacterial strains *escherichia coli* as well as *klebsiella* sp, also two Gram positive bacterial strains *Staphylococcus epidermidis* also *Staphylococcus aureus*, as well as one fungi (*Candida albicans*). In Petri dishes, well diffusion method was used. Two wells in the agar

medium were made in a radius of 1mm, and  $15\mu L$  of each extract were added to the corresponded wells. The plates were incubated at 37 °C for one day, and the inhibition zones were determined.

# 3. Results and Discussion

#### 3.1. Characterization of the plant extract

Table 1 contains the results of qualitative analyses of phytochemicals in peppermint extract. The peppermint extract has shown the presence of carbohydrates, this was in agreement with Petkova *et al.* (2017) who have reported the presence of fructose and sucrose in the extract of peppermint [17]. Phenolic compounds and flavonoids were observed in the extract of peppermint leaves. This content of peppermint results in its strong antioxidant behaviors. This was in agreement with several studies in different countries [18-21]. Proteins, steroid, alkaloids were positively obtained in the extract of peppermint leaves, and this was in agreement with Sujana *et al.* (2013) [22]. On the other hand, saponins were not detected in the extract.

| Test                      | Reagent                 | Color                       | Result   |
|---------------------------|-------------------------|-----------------------------|----------|
| Carbohydrates             | Molish test             | Violet ring                 | Positive |
| Carbonyurates             | Benedict test           | Orange                      | Positive |
| Proteins                  | Biuret test             | Purple blue                 | Positive |
| Steroids                  | Liebermann              | Yellow                      | Positive |
| Steroids                  | Burchard test           | Yellow                      | Positive |
| Phenolic compounds        | Ferric chloride<br>test | Green                       | Positive |
| Tannins and<br>Flavonoids | Lead acetate            | Light yellow                | Positive |
| Alkaloids                 | Mayer's reagent         | White                       | Positive |
|                           | Wagner reagent          | Brown                       | Positive |
| Saponins                  | Fast stirring           | Dense foam for<br>long time | Negative |

Table 1: Qualitative phytochemical outcomes.

Table 2 contains the concentrations of Cu, Zn and Se that obtained by using a flame AAS on peppermint sample. The sample has shown to consist of  $1.16 \ \mu g/L$  of Cu,  $69.36 \ \mu g/L$  of Zn and  $1.26 \ \mu g/L$  Se. The results were in agreement with a previous studies [23, 24]. Zn and Se are introduced in two antioxidant enzymes [25], this high content of these two trace elements would result in the enhancement of the plant's antioxidant capacity.

| Element | Concentration (µg/L) |  |  |
|---------|----------------------|--|--|
| Cu      | 1.16421              |  |  |
| Zn      | 69.3580              |  |  |
| Se      | 1.2576               |  |  |

Table 2: Trace elements in peppermint extract.

#### 3.2. In vitro antimicrobial

Table 3 contains the inhibition zones created by the presence of water or methanol extract of peppermint leaves. The extract solution of peppermint has shown to have an antimicrobial

effect against all of the strains that have been used in this study. The observations has indicated that methanol extract of peppermint exhibited a more powerful antimicrobial effect than the water extract solution. This data were close to the antimicrobial activity of peppermint extract from a previous studies [1, 14, 22, 26-28]. This effect of the peppermint extract could attributed to the high content of medicinal phytochemical such as flavonoids and phenolic compounds whose have an anti-inflammatory characteristics [29].

| Type of microbe  | Water extract | Methanol extract |
|------------------|---------------|------------------|
| E. coli          | 4.0           | 4.5              |
| klebsiella sp.   | 4.2           | 5.1              |
| S. epidermidis   | 3.6           | 3.8              |
| S. aureus        | 4.3           | 5.0              |
| Candida albicans | 2.5           | 3.1              |

| Table 3: Inhibition zones of water and methanol extracts of | peppermints. |
|---|--------------|
|---|--------------|

# 4. Conclusions

The extraction of peppermint leaves have shown the presence of multiple important and medicinal phytochemicals such as the flavonoids and phenolic compounds. Furthermore, it has shown a good percentage of the essential trace elements Cu, Zn, and Se. These components of the peppermint extract would result in enhancement the properties of medical application. The antimicrobial examination of the peppermint extract revealed a good inhibition zones against the five experimented strains.

# References

- 1. Pramila, D., et al., Phytochemical analysis and antimicrobial potential of methanolic leaf extract of peppermint (Mentha piperita: Lamiaceae). Journal of Medicinal Plants Research, 2012. 6(2): p. 331-335.
- Yazdani, D., A. Jamshidi, and F. Mojab, Comparison on menthol content of cultivated peppermint at different regions of Iran. Journal of Medicinal Plants, 2002. 1(3): p. 73-77.
- 3. Dulebohn, J. and R. Carlotti, Soy milk-juice beverage. PCT International Application, WO0249459, 2002. 20.
- 4. Harada, S., H. Ohara, and O. Nishimura, Method for modification of mentha essential oil. Japanese Kokai Tokkyo Koho, JP, 2002. 2002038187(6).
- 5. Güntert, M., et al., Flavor chemistry of peppermint oil (Mentha piperita L.). 2001, ACS Publications.
- 6. Ribeiro, M., et al., Supercritical CO~ 2 Extraction of Peppermint. Influence of Extraction Conditions on the Antioxidant Activity of the Residues. Informacion Tecnologica, 2002. 13(3): p. 185-190.
- 7. Ljubojevic, S. Antioxidative activity of ethanol extract of mint. 2000. sage, vitamin E and synthetic antioxidant BHT. In: International Symposium ....
- 8. Arakawa, T. and K. Osawa, Pharmacological study and application to food of mint flavor-antibacterial and antiallergic principles. Aroma Res, 2000. 1: p. 20-23.
- 9. Pittler, M. and E. Ernst, Peppermint oil for irritable bowel syndrome: a critical review and metaanalysis. The American journal of gastroenterology, 1998. 93(7): p. 1131-1135.
- 10. Nair, B., Final report on the safety assessment of Mentha Piperita (Peppermint) Oil, Mentha Piperita (Peppermint) Leaf Extract, Mentha Piperita (Peppermint) Leaf, and

Mentha Piperita (Peppermint) Leaf Water. International journal of toxicology, 2001. 20 Suppl 3: p. 61-73.

- 11. Sroka, Z., I. Fecka, and W. Cisowski, Antiradical and anti-H2O2 properties of polyphenolic compounds from an aqueous peppermint extract. Zeitschrift für Naturforschung C, 2005. 60(11-12): p. 826-832.
- 12. Hussain, A.I., et al., Seasonal variation in content, chemical composition and antimicrobial and cytotoxic activities of essential oils from four Mentha species. Journal of the Science of Food and Agriculture, 2010. 90(11): p. 1827-1836.
- 13. Rheima, A., Anber, A. A., Shakir, A., Salah Hammed, A., & Hameed, S. (2020). Novel method to synthesis nickel oxide nanoparticles for antibacterial activity. *Iranian Journal of Physics Research*, 20(3), 51-55.
- Mahboubi, M. and N. Kazempour, Chemical composition and antimicrobial activity of peppermint (Mentha piperita L.) Essential oil. Songklanakarin J. Sci. Technol, 2014. 36(1): p. 83-87.
- 15. Mahdi, M.A., et al., Phytochemical content and anti-oxidant activity of hylocereus undatusand study of toxicity and the ability of wound treatment. Plant Archives, 2018. 18(2): p. 2672-2680.
- 16. Tee, E.-S. and C.-L. Lim, Carotenoid composition and content of Malaysian vegetables and fruits by the AOAC and HPLC methods. Food chemistry, 1991. 41(3): p. 309-339.
- 17. Petkova, N., et al., Antioxidants and carbohydrate content in infusions and microwave extracts from eight medicinal plants. Journal of Applied Pharmaceutical Science, 2017. 7(10): p. 055-061.
- Benabdallah, A., et al., Total phenolic content and antioxidant activity of six wild Mentha species (Lamiaceae) from northeast of Algeria. Asian Pacific journal of tropical biomedicine, 2016. 6(9): p. 760-766.
- 19. Uribe, E., et al., Assessment of vacuum-dried peppermint (Mentha piperita L.) as a source of natural antioxidants. Food chemistry, 2016. 190: p. 559-565.
- Rahimi, Y., A. Taleei, and M. Ranjbar, Long-term water deficit modulates antioxidant capacity of peppermint (Mentha piperita L.). Scientia Horticulturae, 2018. 237: p. 36-43.
- 21. Atanassova, M., S. Georgieva, and K. Ivancheva, Total phenolic and total flavonoid contents, antioxidant capacity and biological contaminants in medicinal herbs. Journal of the University of Chemical Technology & Metallurgy, 2011. 46(1).
- 22. Mohammed, S. H., Rheima, A., Al-jaafari, F., & Al-Marjani, M. F. (2022). Greensynthesis of Platinum Nanoparticles using Olive Leaves Extracts and its Effect on Aspartate Aminotransferase Activity. *Egyptian Journal of Chemistry*, 65(4), 1-2.
- 23. Pytlakowska, K., et al., Multi-element analysis of mineral and trace elements in medicinal herbs and their infusions. Food Chemistry, 2012. 135(2): p. 494-501.
- 24. Rheima, A. M., Anber, A. A., Abdullah, H. I., & Ismail, A. H. (2021). Synthesis of Alpha-Gamma Aluminum Oxide Nanocomposite via Electrochemical Method for Antibacterial Activity. *Nano Biomed. Eng*, *13*(1), 1-5.
- 25. Horváth, M. and L. Babinszky, Impact of selected antioxidant vitamins (Vitamin A, E and C) and micro minerals (Zn, Se) on the antioxidant status and performance under high environmental temperature in poultry. A review. Acta Agriculturae Scandinavica, Section A—Animal Science, 2018. 68(3): p. 152-160.
- 26. Mahdi, A., Abbas, Z. S., Hassanain, K., & d Ha, I. (2020). Synthesis, characterization, spectroscopic, and biological activity studies of Nano scale Zn (II), Mn (II) and Fe (II) theophylline complexes.
- Liang, R., et al., Physical and Antimicrobial Properties of Peppermint Oil Nanoemulsions. Journal of Agricultural and Food Chemistry, 2012. 60(30): p. 7548-7555.

- 28. Ohtsu, N., et al., Utilization of the Japanese peppermint herbal water byproduct of steam distillation as an antimicrobial agent. Journal of Oleo Science, 2018. 67(10): p. 1227-1233.
- 29. de Sousa Araújo, T.A., et al., A new approach to study medicinal plants with tannins and flavonoids contents from the local knowledge. Journal of ethnopharmacology, 2008. 120(1): p. 72-80.