Effect of Biological Biostimulator on Green bean Root Biomass

Hassane tahiri¹, Mohammed El yachioui¹, Abderrazzak khadmaoui¹

¹ Faculty of science, Department of biology, Ibn Tofail University, 14000 Kenitra, Morocco

Abstract. The world population continues to grow, there is an urgent need to intensify efforts and innovate to sustainably increase agricultural production. Legumes such as green beans are an excellent source of vegetable protein. In Morocco, the cultivated area of green beans increased from 6,000 hectares in 2017 to 8,000 hectares in 2020, while agricultural production reached more than 200,000 tonnes. This is due to the integration of new agricultural strategies and the use of new monitoring programs. The use of biostimulants as organic fertilizers is proposed to improve crop quality and yield while ensuring greater sustainability of green bean cropping systems. To this end, we proposed to study the efficacy of a biological rooting biostimulant. The results indicate an average increase of 57.32% in root weight from the 90-day growth control and a 59.19% improvement in root length from the 90-day control. This biostimulant shows a very original efficacy in the development of the root system of the green bean plant

 ${\rm Keywords}$: biomass, biostimulant, green bean, integration, rendemant, root sustainability .

1 Introduction

The aim of the new strategies of modern agriculture is to seek sustainable ecological ways to reduce crop dependence on chemical fertilizers [1] and to ensure better yield and quality of food and crops. These chemical fertilizers are non-economic and damaging to the environment and human health [2]. Moreover, the production of organic fertilizer reagents has become a major necessity of agriculture. A biostimulant is a fertilizer that stimulates plant nutrition processes independently of the nutrients. They contain for the sole purpose of improving the agronomic or rhizospheric characteristics of the plant [3]. The global marketplace for plant biostimulants reached \$2 billion in 2019 [4]. However, an estimated 1.5 million hectares of land are at risk and by 2050, 50 percent of arable land is expected to be lost [5]. In recent years, the use of biostimulants as organic fertilizers has become a priority to improve both crop quality and yields while ensuring better sustainability [6, 7]. In addition, biostimulants are applied to directly treat seeds, leaves and other aerial organs as well as soil preparation [8]. In addition, the action of biostimulants is associated with different factors such as: environmental conditions, genetic factor, soil nature and others [9, 10].

In Morocco, the cultivation of pulses occupies a primordial place thanks to their agroeconomic and environmental interests. Taking into account the current socio-economic context, the need to maintain or even increase production of agricultural and agri-food products, in order to be able to meet the demands that are increasing on these products, is currently, imposed. Primary sources of biostimulants include humic acid, fulvic acid, protein hydrolysates, algae extracts, chitosan, inorganic compounds, beneficial fungi and beneficial bacteria [11,12]. The present work is directed towards the establishment of a treatment based on the use of biostimulants and biofertilizers from a biological process of treatment of agrifood by-products and their valorization in the agricultural field, in green beans (*Phaseolus vulgaris*).

2 Material and methods

2.1 Field Preparation

• Soil analysis

The soil of the experimental plot is Clay (45%) -limino (20%)-sandy (35%) with no physical constraints for the cultivation of green beans. We have chosen as parameters pH, nitrogen, matter organic, potassium, phosphorus.

• Soil preparation

For the preparation of the seed bed, a tillage of 25 to 30 cm was carried out using the toothed tools followed by two cross-passes with the cover crop in order to break the clods, aerate the soil and ensure the thickest layer of loose soil with weed removal,

• Type of irrigation

The irrigation system adopted is the drip irrigation sheath, with three irrigations per week of a flow of 2 L/h.

2.2 Plant materials

Our choice has been on the cultivation of green beans (Phaseolus vulgaris), belonging to the family of legumes whose part consumed is the unripe pod, because of its richness in protein, vitamins A and C and mineral salts Ca, Mg, P and K. Sowing was done in March in the field.

2.3 Biostimulants

Rooting Biostimulant: it contains extracts of fermented and stabilized products, rich in free amino acids from hydrolysate obtained by the transformation and stabilization of by-products. However, the presence of polysaccharides, phosphorus and other essential trace elements in combination with amino acids allows having a well-developed root system. It is applied drip every 7 days at a rate of 5 litres/ha. The composition (indicated on the packaging of the product) is as follows:

Extract Algae (30%); Carbohydrate Matter (20%); Total Nitrogen (12%); Ammoniacal Nitrogen (5%); Phosphorus Anhydrous (P2O5) (5%); Potassium Oxide (K2O) (15%); Total Amino Acids (5%); Zinc (0.5%); Manganese (0.5%); Boron (0.35%) and Molybdenum (0.2%).

2.4 Statistical measurement and analysis

Monitoring and evaluation of the effect of biostimulants throughout the culture cycle was carried out on measurements every 15 days using 10 samples per sampling. The parameters taken into consideration for the roots are their weight and length.

The SPSS software carried out the statistical analysis. A descriptive analysis based on the SD mean and a comparison of the mean by the t student test.

3 Results

3.1 Physicochemical and microelement analysis

Table 1 presents the results of the physical-chemical and microelement analysis of the soil selected for the study. The parameters showing very high measurements (exceeding the range) are the basic trend PH (8), Oxidizable organic matter (3.36%), Phosphorus available

Olsen (77 mg/kg), Calcium available meq/100 g (22.1g), Magnesium available meq/100g (5.34g), Potassium available meq/100g (0.99) and Manganese mg/kg (9.34 mg/kg) on the other hand low levels were recorded for Active Limestone %CaCO3, Iron mg/kg and Zinc mg/kg.

Parameters	Results	Standard
pH	8	Very high
Electrical conductivity Us/cm at 20°C	286	normal
Oxidizable organic matter %	3,36	Very high
Nitrogen Dumas mg/kg	2,18	haut
Phosphorus available Olsen mg/kg	77	Very high
Active limestone %CaCO3	2	stockings
Available calcium meq/100 g	22,1	Very high
Magnesium available meq/100g	5,34	Very high
Potassium available meq/100g	0,99	Very high
Available sodium meq/100g	0,76	Haut
Boron mg/kg	0,90	Normal
Iron mg/kg	<4	Low
Manganese mg/kg	9,34	Very high
Copper mg/kg	0,94	Normal
Zinc mg/kg	0,91	Low

Table 1	l∙ Phu	vsicoch	emical	charact	eristics	and	soil	microel	ements
I doite	1 . I II y	Sieden	unical	cinaraci	Clistics	unu	5011	meroer	emento

3.2 Rooting biostimulant effect on root biomass

Table 2 shows the results of the comparison of the mean root weights between the control and the treated over time. In addition, the differences between the mean control and treaty weights appear to be highly significant for all time intervals (p<0.000). Figure 1 presents the results of the evolution in the weight of the green bean root biomass during its vegetative cycle. This evolution appears to increase with time in the control group with a correlation coefficient of (r=0.99; p<0.000) and in the biostimulant group (r=0.99; p<0.000). In fact, the root weights at 15 days of growth in the control and the treated are respectively 1.002 ± 0.02 g and 1.494 ± 0.02 g, this difference between the two average weights was very highly significant (t=-59.278; p<0.000) After 90 days of growth, the control and the treated weights are 20.954 ± 0.06 g and 32.969 ± 0.15 g respectively. Moreover, a marked improvement in root weight is observed under the effect of this biostimulants. Figures 1a and 1b show the density of roots in the green bean plant.

days	Group	Mean±SD	Test levene	p-value	Test	p-value
		Average root			« student »	
		weight				
J 15	control plant	1,002±0,02	0,32	0,578	-59,278	p<0,000***
	treated plant	1,494±0,02				
J30	control plant	1,5±0,01	2,09	0,16	-635,05	p<0,000***
	treated plant	8,508±0,03				
J45	control plant	8,05±0,10	0,59	0,45	-119,83	p<0,000***
	treated plant	14,98±0,15				
J 60	control plant	12,99±0,11	0	1	-203,59	p<0,000***
	treated plant	23,01±0,11				
J 75	control plant	17±0,11	0,76	0,39	-238,23	p<0,000***
	treated plant	28,958±0,12				
J 90	control plant	20,954±0,06	2,11	0,16	-234,46	p<0,000***
	treated plant	32,969±0,15				

Table 2 Comparison of control green bean root weight averages and Time Treated by the student test

J: day; N: effective; Mean: average; SD: standard deviation; Levene test on the equality of variances; Test "t" for equal averages and ***: very highly significant difference.

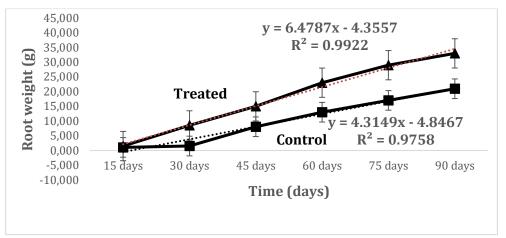
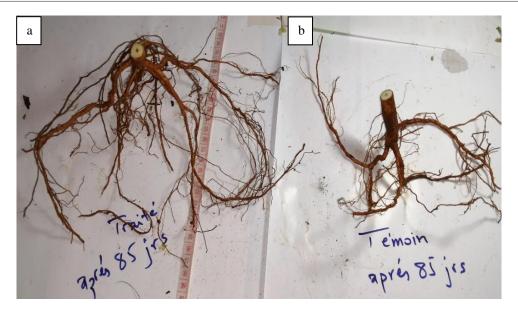


Figure 1 Evolution of the weight of green bean root biomass during its vegetative cycle



3.3 Rooting biostimulant effect on root elongation

The results of the comparison of the average length of the roots are presented in Table 3. There is a very significant difference between the mean length of the control and the treatment as a function of time, and this is apparent during all the sampling intervals of measure (p<0.000). Figure 2 shows the results of the evolution of the length of the green bean root during its vegetative cycle. This evolution appears to increase with time in the control group with a correlation coefficient of (r=0.98; p<0.000) and in the biostimulant group (r=0.97; p<0.000). Indeed, in the control the average length of the roots increases from 5.03 ± 0.05 cm on the 15th day of growth to 30.80 ± 0.05 cm on the 90th day to 49.03 ± 0.05 cm on the 90th day. In addition, the mean treatment length significantly exceeds that of the control (see photo a and b).

Days	Group	Mean±SD	Test levene	p-value	Test	p-value
		Average root length			« student »	
J 15	Témoin	5,03±0,05	0,004	0,950	-177,419	p<0,000***
	Traité	9,03±0,05				
J30	Témoin	8,03±0,06	0,900	0,355	-499,69	p<0,000***
	Traité	21,02±0,04				
J45	Témoin	12,01±0,03	0,002	0,982	-1171,23	p<0,000***
	Traité	30,00±0,03				
J 60	Témoin	19,03±0,05	1,51	0,238	-526,68	p<0,000***
	Traité	37,03±0,09				
J 75	Témoin	26,02±0,04	1,65	0,24	-866,36	p<0,000***
	Traité	45,05±0,05				

Table 3 comparison of root-length means of the green bean between the control and the treated over time by the "student" test

J 90	Témoin	30,80±0,05	1,11	0,16	-102,89	p<0,000***
	Traité	49,03±0,05				

J: day; N: effective; Mean: average; SD: standard deviation; Levene test on the equality of variances; Test "t" for equal averages and ***: very highly significant difference.

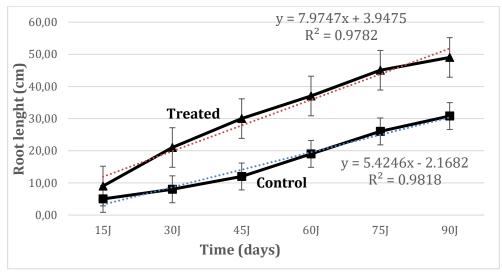
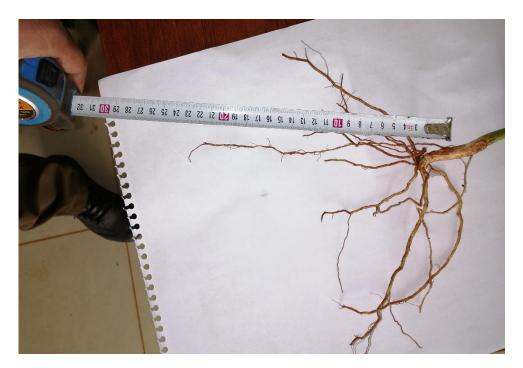


Figure 2 Evolution of the length of green bean roots during its vegetative cycled



4. Discussion

In the face of environmental constraints, plants are able to modify their root systems, rhizosphere, either directly or indirectly by establishing links with other soil organisms [13. Improving biofertilizers has become a priority to meet increased demand for organic products. There are formulations of these biostimulators that can promote and sensitize plant defences and resistance against various environmental stresses. In our study, we conducted field experiments to study the effect of a rooting biostimulant. Results show a significant increase in root weight per plant. These results are consistent with previous work on other biostimulants (humic acid, fulvic acids) Specifying that the perception at the root level of a nutrient concentration in the medium gives rise to the induction of signals leading to local changes in root and foliar architecture. These localized responses can result in increases in root growth and root diameter as well as increases in secondary root numbers [14, 15]. This improvement is mainly due to algae extracts that improve nutrient uptake. In particular, they allow the plant to better tolerate nutrient deficiencies in nitrogen by promoting the expression and/or activity of nitrate reductase through certain compounds (mannitol) [16,17]. The expression of root phosphatases involved in phosphate absorption can also be stimulated by certain algae extracts [18]. Work of [19] showed that the use of Jatropha curcas extracts or powder as a bio-stimulator on green bean crops affects crop growth and yield and showed that the leaves of J. curcas would contain nitrogen and mineral salts that have improved green bean growth and yield. Our results are comparable to those obtained by [20]on the development of green beans treated with liquid extracts of algae. They can be explained by the fact that aqueous extracts of algae contain water-soluble substances such as polysaccharides, phytohormones such as auxin minerals [21] these produce better overall plant growth and yield. Several biostimulants have been used that improve vegetative growth as in chilli [22, 23]

5. Conclusion

The beneficial effect of applying biostimulants can be explained by the direct action of biostimulant components on plant nutrition by acting either on the plant or on the soil. The results obtained during this work showed a significant evolution of treated green beans compared to the control especially the root biomass.

6. References

1. Xu L, D.Geelen, Developing biostimulants from agro-food and industrial by-products. Front. Plant Sci, 9 (2018)

2. M. Dookie, O. Ali, A. Ramsubhag, J. Jayaraman J. Flowering gene regulation in tomato plants treated with brown seaweed extracts. Sci. Hortic, 276:109715(2021);. doi: 10.1016/j.scienta.2020.109715

3. Y. Ole, A.A. Lubyanov, A.Y. Ildus, H.B/ Patrick, Biostimulants in Plant Science: A Global Perspective. Front Plant Sci .;7:2049(2017). doi: 10.3389/fpls.2016.02049.

4. W.Dunham, M. Trimmer.). DunhamTrimmer® global biostimulant report. Section 7: Market overview.60–67(2020°. https://dunhamtrimmer.com/products/biostimulant-globalmarket-report/ 5. Anthony Young. Is there Really Spare Land? A Critique of Estimates of Available Cultivable Land in Developing Countries. Environment, Development and Sustainability.;1:3–18(1999).

6. J.De Saeger, S. Van Praet, D. Vereecke, J.Park, S. Jacques, T. Han et al.). Toward the molecular understanding of the action mechanism of Ascophyllum nodosum extracts on plants. J. Appl. Phycol.;32:573–597. (2020)

doi: 10.1007/s10811-019-01903-9

7. D. Del Buono. Can biostimulants be used to mitigate the effect of anthropogenic climate change on agriculture? it is time to respond. Sci. Total. Environ.;751:141763(2021).

doi: 10.1016/j.scitotenv.2020.141763

8. M.Drobek, M. Frąc, J. Cybulska . Plant biostimulants: Importance of the quality and yield of horticultural crops and the improvement of plant tolerance to abiotic stress-a review. Agronomy 9 (6): 335. (2019)

doi: 10.3390/agronomy9060335

9. M. Ashour, S.M. Hassan, M.E.Elshobary, G.A.G. Ammar, A. Gaber, W.F.Alsanie, et al.. Impact of commercial seaweed liquid extract (TAM®) biostimulant and its bioactive molecules on growth and antioxidant activities of hot pepper (Capsicum annuum). Plants. 10:1045(2021). doi: 10.3390/plants10061045

 M.C.Della Lucia, G.Bertoldo, C.Broccanello, L Maretto L, Ravi S, Marinello F, et al. (2021). Novel effects of leonardite-based applications on sugar beet. Front. Plant Sci. 2021;
doi: 10.3389/fpls.2021.646025

11. Mchugh DJ. "A guide to the seaweed industry". (Rome: Food and Agriculture Organization of the United Nations).2013.

12. Du Jardin P. Plant biostimulants: Definition, concept, main categories and regulation.

Scientia Horticulturae.2015; 196:3-14.

13. Carof M, Laperche A, Cannavo P, Menasseri S, Godinot O, Julbault M, Manzanares-Dauleux M, Guenon R, Jaffrezic A, Pérès G, Le Cadre E. Valorisation des interactions plante-sol pour la nutrition et la santé des plantes. Innovations Agronomiques. 2018; (69): 71-82.

14. Forde B. & Lorenzo H. The nutritional control of root development. Plant Soil.2001;232: 51-68

DOI:10.1023/A:1010329902165

15. Laëtitia Jannin,. Caractérisation des modifications physiologiques et métaboliques induites chez Brassica napus L. par l'apport d'extraits algaux ou d'acides humiques, Universite de caen basseNormandie/UFR. 2012

16. Durand N, Briand X. et Meyer C., « The effect of marine bioactive substances (N PRO) and exogenous cytokinins on nitrate reductase activity in Arabidopsis thaliana », Physiologia Plantarum. 2003;119(4).

17. Phytoma . Mécanismes d'action de l'extrait d'algue GA7 », La défense des végétaux, 2005 ;585 : 42-44

18. Klarzynski O, Fablet E, Euzen M. et Joubert JM. État des connaissances sur les effets des extraits d'algues sur la physiologie des plantes. Phytoma. 2006 ;597.

19. Diédhiou I. Impacts potentiels de l'introduction de Jatropha curcas L. dans un contexte de variabilité et de changements climatiques : impacts environnementaux, intérêts économiques pour les ménages et communautés rurales. Université de Thiès Sénégal, Réseau Sahel. 2009 :19p.

20. Nelson WR, van Staden J. The effect of seaweed concentrate on the wheat culms. Journal of Plant Physiology .1984;82: 199-200.

21. Crouh U, Smith MT, van Staden J, Lewis MJ, Hoad GV. Identification of auxins in a commercial seaweed concentrates. Journal of Plant Physiology. 1992;139: 590-594

22. Barrajón-Catalán E, Álvarez-Martínez FJ, Borrás F, Pérez D, Herrero N, Ruiz JJ, Micol V. Metabolomic analysis of the effects of a commercial complex biostimulant on pepper crops. Food Chem. 2020; 310: 125-818. [CrossRef]

23. Paul K, Sorrentino M, Lucini L, Rouphael Y, Cardarelli M, Bonini P, Reynaud H, Canaguier R, Trtílek M, Panzarová K, et al. Understanding the biostimulant action of vegetal-derived protein hydrolysates by high-throughput plant phenotyping and metabolomics: A case study on tomato. Front. Plant. Sci. 2019; 10: 1–17. [CrossRef] [PubMed]